



ISSN 2347-2677

IJFBS 2017; 4(2): 47-49

Received: 09-01-2017

Accepted: 10-02-2017

Rumana Shaikh

Department of Zoology, Pemraj
Sarada College, Ahmednagar,
Maharashtra, India

Atul R Chourpagar

Department of Zoology,
Dadapatil Rajale Arts and
Science College, Adinathnagar,
Tal. Pathardi, Dist.
Ahmednagar, Maharashtra,
India

GK Kulkarni

Retd. Professor & Head,
Department of Zoology,
Babasaheb Ambedkar
Marathwada University,
Aurangabad, Maharashtra, India

Biochemical composition of selected Meiobenthic fauna, cultured copepod *Mesocyclops leuckarti* (Claus, 1857) from Mombatta Lake

Rumana Shaikh, Atul R Chourpagar and GK Kulkarni

Abstract

The present study was carried out from the laboratory culture; *Mesocyclops leuckarti* collected from Mombatta Lake at Daulatabad was sieved through 42 µm plankton net. 100 mg of cultured *Mesocyclops leuckarti* was taken for the biochemical estimation. The biochemical composition of *Mesocyclops leuckarti* shown the protein content was $58.33 \pm 0.55\%$ dry weight, lipid was $10.16 \pm 0.10\%$ dry weight, carbohydrates was $18.10 \pm 0.24\%$ dry weight and the water content was $93.25 \pm 0.77\%$.

Keywords: Biochemical composition, copepod, *Mesocyclops leuckarti*, Mombatta Lake

1. Introduction

Sleep The production of planktonic organisms is good nutritional condition to feed fish larvae and fingerlings is a basic requirement in fish culture. Although this procedure ensures a quick response in terms of algal biomass increase, both zooplankton composition and nutritional condition change abruptly, causing low survival rates of fish fingerlings due to bad quality of food (Santeiro and Pinto-Coelho, 2000) [18]. An adequate plankton biochemical composition ensures the nutritional requirements for fish fingerlings especially during their initial development stages. Rajkumar and Vasagam (2006) [14] reported that many copepods are identified as potential live feed organisms and active investigations are carried out.

A wide variety of live organism has been utilized in larvae culture, mainly because of their nutritional value, which is higher than that of prepared diets. Copepods are a well-known natural food source from fish larvae and fingerlings, and the larvae of certain fishes prefer nauplii and copepodids over rotifers (Meeren, 1991; Schipp *et al.*; 1999) [11, 21]. Copepods constitute an important component of the food chain in aquatic ecosystem and accepted to be highly satisfactory for larvae of prawn and fin-fish species. Biochemical studies have shown that copepods are rich in protein, lipids, essential amino acid (EAAs), and essential fatty acids (EFAs) which can provide enhanced reproduction of brood stock, augmented growth, immune stimulation, and colour enhancement in prawns and fishes (Altaff and Chandran 1989; Safiullah, 2001) [1, 17].

Many authors have reported the utilization of copepods from wild and cultured sources from higher yields of prawn in ponds (Goswami *et al.*, 2000; Ashok *et al.*, 2005; Ashok and Rajkumar, 2007; Rajkumar *et al.*, 2008) [7, 2, 3, 15]. Studies on the biochemical estimation of different copepods and their utilization as food for the freshwater post larvae of prawn and fish are limited. Therefore, present study is focused on the biochemical composition viz. protein, lipid, carbohydrate and water content of laboratory cultured copepod *Mesocyclops leuckarti*.

2. Material and methods

The laboratory culture, *Mesocyclops leuckarti* collected from Mombatta Lake at Daulatabad was sieved through 42 µm plankton net. Collected *Mesocyclops leuckarti* was first washed with clean water to remove detritus. It was filtered through filter paper and was dried a blotting sheet. From this blot dried material, 100 mg of cultured *Mesocyclops leuckarti* was taken for the biochemical estimation by adopting Biuret method of Lowry *et al.*, (1951) [9] for protein, Dezwaan and Zandee, (1972) [6] using the anthrone reagent for carbohydrates and the total lipid estimated by the Vanillin reagent method by Barnes *et al.*, (1973) [4]. The water content of *Mesocyclops leuckarti* was determined according to Clegg (1974) [5]. Statistical analysis was performed by Mungikar (2003) [12].

Correspondence

Rumana Shaikh

Department of Zoology, Pemraj
Sarada College, Ahmednagar,
Maharashtra, India

3. Results

The biochemical composition of *Mesocyclops leuckarti* shown in table 1. The protein content was $58.33 \pm 0.55\%$ dry weight, lipid was $10.16 \pm 0.10\%$ dry weight, carbohydrates was $18.10 \pm 0.24\%$ dry weight and the water content was $93.25 \pm 0.77\%$.

Table 1: Showing the biochemical composition of culture copepod, *Mesocyclops leuckarti*.

Parameters	Biochemical composition (%), Dry weight
Protein	58.33 ± 0.55
Lipid	10.16 ± 0.10
Carbohydrates	18.10 ± 0.24
Moisture	93.25 ± 0.77

Mean \pm S.D. of 3 observations

4. Discussion

Perumal *et al.*, (2009) [10] reported the protein content of wild copepods varied from 59.53 to 69.61% in *Oithona similis* and 67.33 to 75.45% in *Acartia spinicauda* from Parangipatti coast. Sipauba Tavares 2001 [22] reported 64% protein content in cultured *Argyrodiaptomus furcatus*. Hossain *et al.*, (2002) [8] while working on diet of mahseer fingerlings reported that feed containing 40% protein exhibited the best growth response while diet containing 20% protein resulted is the poorest growth. The present study show that protein 58.33% is the major chemical constituent of cultured *Mesocyclops leuckarti*.

Santeiro *et al.*, (2006) [19] reported 9.14% lipid content in copepod *Thermocyclops sp.* Rao and Krupanidhi (2001) [16] showed that variation in the lipid content can be attributed to its storage and utilization during period when it serves as an effective energy reserve. The present study 10.16% lipid content are present in cultured *Mesocyclops leuckarti* the lipid content of tropical zooplankton when compared to temperate zooplankton is significantly low which may be due to the hydrological condition and the type of availability of food organisms in environment (Ashok *et al.*, 2005) [2].

The carbohydrates content in cultured copepod *Mesocyclops leuckarti* showed 18.10%. According to Perumal *et al.*, (2009) [13] low carbohydrate content may be attributed to the fact that glycogen is the usual storage carbohydrate in many zooplankton like *Oithona similis* 7.98% and 6.59% in *Acartia spinicauda*. Comparatively *Mesocyclops leuckarti* show that the carbohydrates content a much higher 18.10% than reported.

In the present study, *Mesocyclops leuckarti* the water content was found to be 93.25%. Vengadeshperumal *et al.*, (2010) [18] has reported $82.94 \pm 0.60\%$ water content in another cultured copepod *Acartia centrura*. In cultured copepods *Oithona similis* varied from 79.22 to 83.87% and in *Acartia spinicauda* 82.06 to 85.80% reported by Perumal *et al.*, (2009) [13]. According to Madhupratap *et al.*, (1997) [10] different copepod species have a different protein, lipid and carbohydrates contents.

5. Conclusion

It is important to note that a species of choice could be used to obtain best result in fish culture.

6. Acknowledgement

This study was supported by the University Grant Commission (UGC), under the major research project no. F. 33-328/2007 (SR). Authors are indebted to Prof. G. K. Kulkarni for valuable guidance.

7. Reference

1. Altaf K, Chandran MR. Sex- related biochemical investigation of the diaptomid *Heliodyptomus viduus* (Gurney) (Crustacea: Copepoda). Proc. Indian Sci. Acad. (Animal Sci.). 1989; 98:175-179.
2. Ashok PV, Perumal P, Rajkumar M. Biochemical composition of some marine copepods. Res. J. Chem. Environ. 2005; 9:36-41.
3. Ashok PV, Rajkumar M. Biochemical composition of cultured copepod, *Acartia spinicauda*, Giesbrecht. Aquacult, 2007; 8:219-224.
4. Barnes, Blackstoch J. Estimation of lipids in marine animals and tissues. Detailed investigation of the sulpho-phosphovanillin method for total lipids. J. Exp. Mar. Biol. Ecol. 1973; 12(1):103-108.
5. Clegg JS. The control of emergence and metabolism by external osmotic pressure and the role of free glycerol in developing cysts of *Artemia salina*. J. Exp. Biol. 1974; 41:879-892.
6. De Zwann A, Zandee DI. The utilization of glycogen and accumulation of some intermediate, during anaerobiosis in *Mytilus edulis*. Comp. Biochem. Physiol. 1972; 43(B):47-52.
7. Goswami SC, Krishnakumari L, Shrivastava Y. Diet variation in zooplankton and their biochemical composition from Venguria to Ratnagiri, West Coast, India. J. Mar. Sci. 2000; 29:177-180.
8. Hossain, M.A., Hasan, N., Azad Shah, A.K.M. and Hussain, M.G. Optimum Dietary Protein Requirement of Mahseer, *Tor putitora* (Hamilton) Fingerlings. Asian Fisheries Science Asian Fisheries Society, Manila, Philippines, 2002, 15: 203-214.
9. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurement with the folin-phenol reagent. J. Biol. Chem. 1951; 193:265-275.
10. Madhupratap, M., P. Venugopal and Haridass. Biochemical studies on some tropical estuarine zooplankton. Indian J. Mar. Sci., 1979, 8: 155-157.
11. Meeren, T. V. D. Selective feeding and prediction of food consumption in turbot larvae (*Scophthalmus maximus*) reared on the rotifer *Brachionus plicatilis* and natural zooplankton. Aquaculture, 1991, (93): Pp 35-55.
12. Mungikar aM. An introduction to biostatistics. Saraswati Publ. Aurangabad. 2003, 1-63.
13. Perumal P, Rajkumar M, Santhanam P. Biochemical composition of wild copepods, *Acartia spinicauda* and *Oithona similis*, from Parangipettai coastal waters in relation to environmental parameters. Journal of Environmental Biology November. 2009; 30(6):995-1005.
14. Rajkumar M, Vasagam KP. Suitability of the copepod, *Acartia* clause as a live feed for Sea-bass larvae (*Lates calcarifer* Bloch): Compared to traditional live food organisms with special emphasis on the nutritional value, Aquaculture, 2006; 261:649-658.
15. Rajkumar M, Vasagam KP, Perumal P. Biochemical composition of wild copepods, *Acartia erythroa* Giesbrecht and *Oithona brevicornis*. Giesbrecht, from Coleroon coastal waters, South East coast of India. In: Advances in Aquatic Ecology, (Ed: V. B. Sakhare), Daya Publishing House, New Delhi- 35. 2008; 2:1-20.
16. Rao, N. I. & Krupanidhi, G. Biochemical composition of zooplankton from the Andaman Sea. J. Mar. Biol. 345

- Assess. India. 2001, 43, 49-56.
17. Safiullah A. Biochemical and nutritional evaluation and culture of freshwater live food organisms for aqua hatcheries. Ph.D. Thesis, University of Madras, 2001, 103.
 18. Santeiro RM, Pinto-Coelho RM. Efeitos de fertilizaco na biomassa e qualidade nutricional do zooplankton utilizado para alimentaco de alevinos na estaco de hidrobiologia e piscicultura de Furnas, MG. Acta Sci., Maring, 2000; 22(3):707-716.
 19. Santeiro R.M., Pinto-Coelho R. M., and Sipauba Tavares L. H. Diurnal variation of zooplankton biochemical composition and biomass in plankton production tanks biomass in plankton production tanks, Acta Sci. Biol. Sci. Maring, 2006; 28(2):103-108.
 20. Santiago CB, Lowell RT. Amino acid requirements for growth of Nile tilapia. Nutr. 1988; 118:1540-1546.
 21. Schipp, G. R., Bosmans, J. M. P and Marshall, A. J (1999): A method for hatchery culture of tropical calanoid copepods, *Acartia* spp. Aquaculture, 1999, (174): Pp 81–88.
 22. Sipauba-Tavares, L.H., Bachion, M.A. & Braga, F.M.S. Effects of food quality on growth and biochemical composition of a calanoid copepod, *Argyrodiaptomus furcatus*, and its importance as a natural food source for larvae of two tropical fishes. Hydrobiologia, 2001, 453/454:393-401.
 23. Vengadeshperumal, N., P. Damotharan, M. Rajkumar, P. Perumal, S. Vijayalakshmi and T. Balasubramanian. Laboratory Culture and Biochemical Characterization of the Calanoid Copepod, *Acartia southwelli* Sewell, 1914 and *Acartia centrura* Giesbrecht, 1889. Advan. Biol. Res., 2010, 4 (2): 97-107.