

International Journal of Fauna and Biological Studies

Available online at www.faujournal.com

I
J
F
B
S
International
Journal of
Fauna And
Biological
Studies

ISSN 2347-2677

www.faujournal.com

IJFBS 2019; 6(2): 89-98

Received: 08-01-2019

Accepted: 12-02-2019

Divya Gupta

Ranjan Plant Physiology and
Biochemistry Laboratory,
Department of Botany,
University of Allahabad,
Prayagraj, Uttar Pradesh, India.

Abreeq Fatima

Ranjan Plant Physiology and
Biochemistry Laboratory,
Department of Botany,
University of Allahabad,
Prayagraj, Uttar Pradesh, India.

Sunita Kumari Singh

Government College Rajpur,
Balrampur, Chattisgarh, India.

Pratibha Singh

Ranjan Plant Physiology and
Biochemistry Laboratory,
Department of Botany,
University of Allahabad,
Prayagraj, Uttar Pradesh, India.

Sheo Mohan Prasad

Ranjan Plant Physiology and
Biochemistry Laboratory,
Department of Botany,
University of Allahabad,
Prayagraj, Uttar Pradesh, India.

Correspondence**Pratibha Singh**

Ranjan Plant Physiology and
Biochemistry Laboratory,
Department of Botany,
University of Allahabad,
Prayagraj, Uttar Pradesh, India.

Repercussion of soil pollution on plants

Divya Gupta, Abreeq Fatima, Sunita Kumari Singh, Pratibha Singh and Sheo Mohan Prasad

Abstract

Healthy environment is been established by healthy soil. But nowadays, soil pollution is becoming a major threat to environment, so we need to overcome fast. The soil is the home for a large part of bacterial biodiversity and other microscopic and macroscopic living organisms. Many of the natural processes and destructive human activities lead to the contamination of the soil where heavy metal contamination is the major issue in the environmental problems of soil. Other pollutants mainly include chemicals such as pesticides, lead, ammonia, petroleum hydrocarbons, herbicides, nitrate, naphthalene, mercury etc in an excess amount. These chemicals reduce the soil quality by hampering its fertility and further make the soil inhabitable for macro-organisms and microorganisms existing in the soil. Moreover, food security is also been hindered as soil pollution minimizes the yield and quality of the crops. This review article will brief the status of soil contamination, its origin and risks in the commencement and after effects on plants and their productivity status. Proper action is to be taken as healthy soil is considered non-renewable, precious commodity for sustainable and food protected future.

Keywords: agrochemicals, biodiversity, contamination, food security, heavy metals, soil pollution

Introduction

Pollution affects everything from the air we breathe to the food we eat. It is invisible thereby can pose serious threats and even go untraced. Presence of chemicals, pesticides, fertilizers etc. in the soil above a certain limit causes disturbance in the growth and survival of the plants and animals is known as soil pollution. Soil pollution disturbs the growth of the flora by creating various stresses. Extensive use of pesticides, herbicides and fertilizers causes depletion of soil and later on converts it into desert. Nearly, 20 million hectares of agricultural land is irrigated with untreated waste water (WHO, 2006) ^[101] which is also one among other sources of soil pollution. These pollutants increase the salinity of the soil and make it more redundant for the growth of the plants. This in turn causes deprivation of minerals from the soil and makes it infertile.

World population is increasing day by day whereas the productivity of the crop is not increasing in the same proportion. Pollution seriously affects soil functions, by decreasing the number and diversity of species inhabiting in the same soil. Successive decrease in the microbial activity of soil reduces nutrient cycling and in the consequence of which the soil structure gets affected and modified as well as the fertility for crop production (Jacoby *et al.*, 2017) ^[41]. Different types of pollutant are reported in the soil named as radioactive fallout, microbial pathogenesis, salinity and metal contaminants (FAO, 2009) ^[23]. Other causes include the rupture of underground storage tanks, percolation of contaminated surface water, leaching of wastes from landfills, discharge of industrial effluents into the soil, unfavorable irrigation practices, improper septic management, sewage and fuel leakages from automobiles etc. All these pollutants get washed with rain water and seep into the soil later on causing serious health concerns of plants as well as animals. Therefore, these pollutants need serious concern for their eradication from the agricultural soil. The different pollutants existing in soil and their after effects on plants are well illustrated in Fig.1. Therefore, this review provides a detailed study on the type, occurrence, causes and extent of susceptibility of pollutants.

Heavy Metals

Heavy metals are the kind of major pollutants of soil pollution as they are complex having more perseverance properties to remediate in nature. They also possess metallic properties which sum ups with malleability, conductivity, ductility, ligand specificity and cation

stability. Their atomic number is greater than 20 with relative high atomic weight (Raskin *et al.*, 1994) ^[80]. Some metals namely Co, Fe, Mn, Cu, Mo, Ni, Zn and V are required in lesser amount to the organisms but as their amount is increased due to geologic and anthropogenic activities viz., burning of fossil fuels, mining and smelting of metals, use of fertilizers and pesticides in agriculture, production of batteries and other metal products in industries, sludge, sewage and municipal waste disposal so become considerably harmful for both the plants and animals (Alloway, 1990; Shen *et al.*, 2002) ^[3, 86]. Some other metals are also lined up like Pb, Cd,

Hg and As coming under metalloid are main threats to the plants and animals as they do not have any beneficial effects. In soil, metal exists as separate ion or in mixtures with other metals or with soil constituents. These constituents contain free metal ions, resolvable metal compounds, unresolvable inorganic metal compounds such as carbonates and phosphates in soil solutions, non-exchangeable ions and exchangeable ions which are adsorbed on the faces of inorganic solids and also metals attached to silicate minerals (Marques *et al.*, 2009) ^[61].

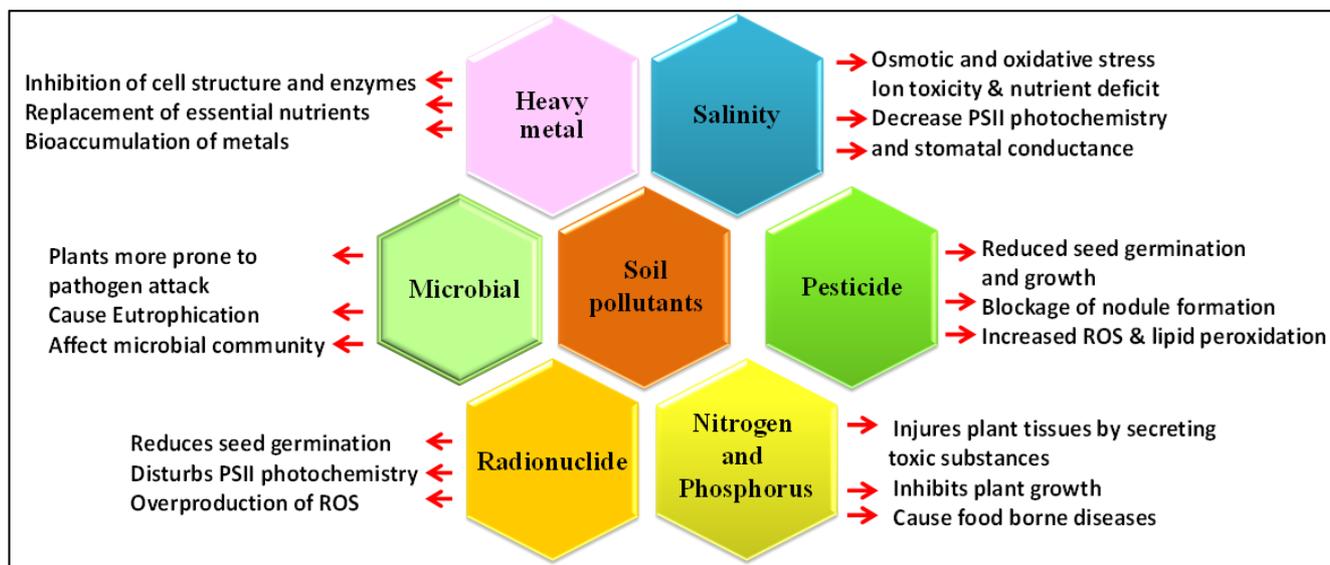


Fig 1: Different types of soil pollutants and its effects on plant physiology and metabolism

A. Heavy Metal Availability in Soil as Affected by Soil Physical Properties

It has also been noted that many of the soil properties affects diversely the availability of metal in the soil. Significantly soil pH, moisture content, water holding capacity are the main properties of soil which have positive or negative correlation with the heavy metals (Sharma and Raju 2013 ^[84]; Haddad *et al.*, 2018 ^[35]; Liu *et al.*, 2018) ^[54]. In accordance to it, in the roots of *Thlaspic aerulescens*, availability of Cd and Zn decreased with increased soil pH (Wang *et al.*, 2006) ^[100]. Also, heavy metal availability is decreased by organic matter and hydrous ferric oxide through the process of immobilization of these metals (Yi *et al.*, 2007) ^[103]. Conversely, heavy metals are known to modify the biological properties of soil (Friedlova, 2010) ^[24].

B. Heavy Metal Affects Biological Properties of Soil

The harmfulness of metal on soil microorganism depends on the various factors such as soil pH, temperature, inorganic anions and cations, organic matter, clay minerals and chemical forms of the metal (Baath, 1989 ^[6]; Friedlova, 2010 ^[24]; Haddad *et al.*, 2018) ^[35]. Many of the studies describe the discrepancies related to heavy metal contributing soil pollution. Some of the researchers recorded positive effects while others recorded negative effects of heavy metal on biological properties of soil (Smejkalova *et al.*, 2003 ^[92]; Castaldi *et al.*, 2004 ^[16]; Friedlova, 2010 ^[24]). If one metal is present in the soil it affects the other one as well. This indicates that an antagonistic and synergistic behavior exist among heavy metals. Salgare and Acharekar (1992) ^[81] reported that in the presence of Cd, the inhibitory effect on the

total amount of mineralized C was antagonized. Likewise, Cu and Zn as well as Ni and Cd have been testified to compete for the same membrane carriers in plants (Clarkson and Luttge, 1989) ^[18]. In contrast, the toxicity of Zn was increased in presence of Cu in spring barley (Luo and Rimmer, 1995) ^[56].

C. Heavy Metal Effects on Plants

Heavy metals which are soluble in soil solution or are solubilized by root exudate are been up-taken by the plants and affects directly as well as indirectly (Blaylock and Huang, 2000). Initially, in limited amount these heavy metals are required for their growth but above the threshold limits are toxic to the plants. Plants have the unique ability to accumulate the essential metals but this also negatively enables them to accumulate the non-essential metals (Djingova and Kuleff, 2000) ^[21]. In plants, the direct toxic effects with increasing concentration of heavy metals pertains to the inhibition in cell structure as well as enzymes present in the cytoplasm mainly due to induced oxidative stress (Assche and Clijster, 1990) ^[5]. The indirect toxic effects include the replacement of essential nutrients at cation exchange sites of plants (Taiz and Zeiger, 2002) ^[96].

Soil with heavy metal concentrations are further characterized by decline in soil nutrients due to decrease in decomposition of organic matter and reduction in the number of soil microorganism. These direct and indirect toxic effects ultimately leads to the destruction in plant growth which further can also lead to the death of the plant. Presence of some non-beneficial heavy metals at very low concentrations in the growth medium has adverse effects on the plants. Kibra

(2008)^[52] noted that soil adulterated with 1 mg Hg/kg exerted substantial decline in height of rice plants as well as tiller and panicle construction. Similarly 5mg/L Cd in soil solution led to decline in root and shoot growth in wheat (Ahmad *et al.*, 2012)^[1]. The possible reasons behind the decline in growth parameters in polluted soils might be due to decrease in photosynthetic activities, plant mineral nutrition, and abridged activity of some enzymes (Kabata-Pendias, 2001)^[50]. In contrast to these findings, small concentrations of these heavy metals possibly enhance the growth and development of the plant but as soon as the amount increases in soil, its positive effects get reversed. For instance, many of the researchers examined that low concentration of Co i.e. 50 mg Co/kg increase the nutrient and biochemical content, antioxidant enzyme activities while it had devastating effects on nutrient and biochemical content, antioxidant enzyme activities in plant growth with high concentrations at 100 mg Co/ kg to 250mg Co/ kg when compared to the untreated seedlings of tomato plant (Jayakumar *et al.*, 2007)^[47]; Jayakumar *et al.*, 2008^[46]; Jayakumar *et al.*, 2013)^[48]. Similarly, Zn at concentration of 25 mg/L in the soil solution improved the growth and physiology of cluster beans. On contrary, 50mg Zn/L of soil solution was noted to result in starting of reduction in growth and physiology of plants (Manivasagaperumal *et al.*, 2011)^[59].

The sources of metal toxicity are so diverse that at times there is more than one heavy metal in the soil which affects plant toxicity in both antagonistic and synergistic relationship. One of such study was reported by Nicholls and Mal (2003)^[69] where both the heavy metal Pb and Cu at their high (1000 mg/kg each) and low concentrations (500 mg/kg) damaged vigorously the stem and leaves of *Lythrum salicaria* and ultimately lead to death. The authors analysed that there was no synergistic relationship between the heavy metals probably because the concentrations were so high to have interactive relationship among them. Ghani (2010)^[30] tested the effect of six heavy metals (Cd, Cr, Co, Mn, Hg and Pb) on the growth and physiology of maize and showed existence of antagonistic relationship among the heavy metals where growth and protein content of maize seedlings was drastically reduced. The researchers were amazed on seeing the combined effect of two heavy metals as detrimental as the effect of most harmful heavy metal. The range of toxicity of these metals was in the following order: Cd > Co > Hg > Mn > Pb > Cr.

Plants also possess variety of mechanisms to avoid the toxic effects caused by heavy metals in soil. Some plants are able to tolerate the high concentrations of heavy metals in the surroundings. Baker (1981) reported three supposed mechanisms acquired by these plants to tolerate these heavy metals namely (i) exclusion- the act of not allowing metal transport to take part in an activity and conservation of a persistent metal amount in the shoot over a extensive range of soil concentrations; (ii) inclusion- maintaining the linear relationship of metal concentrations in the shoot as well as in the soil solution and (iii) Bioaccumulation- accumulation of metal of both high and low concentration in soil at a rate faster than that at which it is lost by catabolism and excretion in shoot and root of the plants.

Salinity

A. Causes and Distribution

With the onset of 21st century, it was marked largely as environmental pollution basically due to increased

salinization of soil and water. Among the other environmental stressors, soil salinity is found to be most devastating in terms of destruction in cultivated land area and crop productivity and quality. With the increasing human population, there are decreasing available land area for cultivation, consequently, reduction in crop productivity which leads to the major threats, needs to be cope up for stable agricultural sustainability (Shahbaz and Ashraf, 2013)^[83]. Saline soil is explained when the electrical conductivity of the saturation extract at 25 °C in the root sector exceeds 4 dSm⁻¹ (approx. 40 mM NaCl) has an exchangeable sodium of 15% (Munns, 2005; Jamil *et al.*, 2011)^[66, 42]. Factors like low precipitation, high surface evaporation leads to extreme climate, weathering of native rocks, bad irrigation practices with saline water and poor, unaware cultural practices are held to be responsible for salinity.

Worldwide salinized region are growing at a rate of 10% annually, which by the coming year 2050, will become more than 50% of the arable land leads to low agricultural potential (Jamil *et al.*, 2011)^[42]. Naturally, salts are present as ions in the soil. The sources of getting salt in the soil are probably observed to be irrigation with saline water or with fertilizers, getting upward in the soil through shallow groundwater, also get accumulated under low precipitation as not able to leach ions from soil outline and causing soil salinity (Isayenkov and Maathuis, 2018)^[40].

In India, saline soil covers approx. 7 million hectares of land (Patel *et al.*, 2011)^[76]. Most of the rejoin include indo-gangetic plane covering adjoining states like Uttar Pradesh, Bihar, Haryana and Punjab and some parts of Rajasthan. Some of the arid and semi-arid regions also get affected by saline soil which includes Gujarat, Madhya Pradesh, Maharashtra and Karnataka.

B. Effects of Salinity

Salinity disturbs the ecological balance of that region as it not only decline the agricultural productivity of crops but also have a major impact on the physico-chemical properties of soil mainly causing the soil erosion (Rahneshan *et al.*, 2018)^[78]. It effects seed germination, nutrient and water uptake, plant growth and reproductive development (Akbarimoghaddam *et al.*, 2011)^[2], causes physiological damages as ion toxicity, nutrient deficiency, osmotic, metabolic and oxidative stress, when the salt accumulation in the soil are higher than inside the root cells, the soil will draw water from the root, and the plant will wilt and die (Singh *et al.*, 2019)^[94]. Also, the salinity causing acidity and alkalinity of the soil due to the imbalance in pH value as a result of excess sodium may end to micronutrient scarcities (Rahneshan *et al.*, 2018)^[78]. With reference to salinity, plants sensitivity varies among different species as tomatoes, onions and lettuce comes under the resistant variety and on contrary, halophytes are the tolerant variety. Phosphorus uptake was considerably declined as ions of phosphates get precipitated with Ca ions (Bano and Fatima, 2009)^[8]. One of the most important physiological parameter is the photosynthesis which is also damaged on account of salinity along decline in leaf area, chlorophyll content, transpiration rate and stomatal conductance and also to some extent affects the PSII photochemistry and electron transport chain (Parihar *et al.*, 2015)^[73]. It is also reported to result in impairment in the supply of photosynthetic assimilate and the regarding hormones (Ashraf and Harris, 2004^[4]; Netondo *et al.*, 2004)

[68]. Plant growth development is also hampered by inhibition at the level of microsporogenesis and elongation of stamen filament during reproductive stages resulting in the enhanced ovule abortion, programmed cell death and senescence of fertilized embryos. All these factors starting from the germination to the reproductive level and finally ending towards senescence result from osmotic stress at the physiological, biochemical level and at the molecular level (Parihar *et al.*, 2015 [73]; Isayenkov and Maathuis, 2018) [40]. At the molecular level, ion toxicity has been portrayed by the replacement of K^+ by Na^+ in biochemical reactions, and Na^+ and Cl^- ions persuaded conformational deviations in proteins. Biologically, inside the cell is K^+ ion which is important and can't be replaced by any another ion. In the translation process a sufficient amount of K^+ ion is needed for the binding of tRNA to ribosomes for protein synthesis (Zhu 2002) [107]. Pruning in spikelets per spike, stunted spike emergence and reduced fertility all at the reproductive phase was found to be more profound in the wheat seedlings exposed to 100-175 mM NaCl ultimately resulting in poor grain (Munns and Rawson 1999) [67]. So, such combined effects of salinity also damage the cell cycle and cell differentiation. Recent report confirms the effect of salinity on RNA, DNA as well as mitosis, meiosis, protein and enzymatic activity (Tabur and Demir 2010; Javid *et al.* 2011) [95, 45].

Pesticides

Pest considered an important aspect for the farmers in agriculture in both developed and developing countries. Agrochemicals were introduced to the agriculture decades ago aiming to enhance crop productivity, yield and also to protect the vegetation from weeds and pest. It has also been estimated that usable amount of pesticides which is actually used to control pest and reaches the action site is only a small part (<0.1%), and rest of the big portion is lost via run-off, foliar spray drift, photo degradation, off-target deposition (Silva *et al.*, 2019) [87]. However, Joy *et al.* (2013) [49] reported that in sorghum translocation in plant and adsorption of pesticides by the soil is not justified to gain resistance. As pesticides have high persistence in the soil so in agriculture unselective and unconcerned use of these chemicals have major associated damages such as environmental pollution, pesticides residue in food, fruits, fodder, vegetables, soil etc and most importantly causing ecological imbalance and pest resurgence etc. These harmful effects are not only the outcome of chemically active ingredients and their allied impurities but also due to the solvents, emulsifiers, carriers and other elements of the articulated product. The survey created by United Nations Environment Protection in 1972 detailed that nine of the 12 most undesirable persistent organic pollutants are pesticides which are used in agriculture crop since long time and for public health related vector control program.

Pesticides are known to create damages at all stages of plant life, starting from seed germination. Rajashekar *et al.* (2012) [79] analyzed that with the increasing dose of pendamethalin in *Zea mays* L. NAAC-6002, seed germination rate reduced drastically to 69% in treated seedlings. Similarly, length of plumule decreased by 77% and the length of radical decreased up to 90% at 10 ppm. Dubey and Fulekar (2011) [22] reported that the use of chlorpyrifos, cypermethrin and fenvalerate at amount of 0-100 mgKg⁻¹ through spiked soil caused significant decrease in seed germination. But at higher concentrations (75 and 100 mg/kg) of chlorpyrifos, this

decrease was more profound compared to cypermethrin and fenvalerate in *Cenchrus setigerus* Vahl, *Pennisetum pedicellatum* Tan seedlings. Moore and Kroger (2010) studied the combined and single treatment effect of three insecticides and two herbicides, asfipronil showed lowest 76% seed germination, but diazinon attested well in germination (85%) which is more than the control. Among herbicides combined treatment of atrazine and metalachlor was found to have better germination rate by 85% than from the single application of atrazine which was 72% in rice seedlings. Basantani *et al.* (2011) [10] reported that the percentage of growth, root length and shoot length declined with the application of glyphosate (10mM) in two varieties of *Vigna radiata*. Similar observations were reported by Singh and Prasad (2018) [90] while studying the effect of chlorpyrifos, dimethoate and dieldrin applied twice at its recommended dose on the growth parameters of *Spinacia oleracea*. It was explained on the basis of decreased photosynthetic pigments and increase in reactive oxygen species and oxidative damages in turn. Chlorotoluron obstructed the photosynthetic electron transport in higher plants (Fuerst and Norman 1991) [27] and interrupted PSII reaction centre (Barry *et al.* 1990). Parween *et al.* (2011) [75] reported that insecticide chlorpyrifos significantly increased the plant height, total leaf area, number of branches, leaf per plant and plant biomass when applied at 0-1.5mM but as the concentration increases it had all negative impact on above parameters in *Vigna radiata* L seedlings. *Cicer arietinum* L. was severely affected by some of the insecticides such as lorsban, decis, pyrifos, karate, ripcord when applied at levels 875 mL acre⁻¹, 200 mL acre⁻¹, 1125 mL acre⁻¹, 250 mL acre⁻¹, 225 mL acre⁻¹ respectively at pod initiation stage and continuously sprayed for 45 days after planting. After pyrifos treatment nodulation was significantly blocked. Contrary to it, grain vintage was significantly greater as compared to other insecticide (Mahmood and Shah 2003; Khan *et al.*, 2009) [57, 51]. Fungicide captan at higher concentration showed decline in photosynthetic pigments (chlorophyll *a*, *b* and carotenoids) but at lower concentration or recommended doses it increased the photosynthetic pigments in pepper leaves (Tort and Turkyilmaz, 2003). Mishra *et al.* (2008) [98, 63] reported that, photosynthetic oxygen yield, photosystem II and its chemistry, CO₂ fixation along with whole chain activities were found to decline adversely at all doses of dimethoate. Suri and Singh (2011) [94] found that after treatment of *Oryza sativa* L. seedlings with quinalphos, chlorpyrifos, methyl parathion, endosulfan, imidacloprid, and deltamethrin for three days at the interval of 10 days in the potted plant with half of the recommended doses revealed that sugar, protein and amino acid reduced in content. The amount of total phenols in leaf sheaths and blades of methyl parathion, deltamethrin, and quinalphos treated plants of the two varieties got lowered. On the other hand, chlorpyrifos and endosulfan significantly lowered or did not influence the content of reducing sugars in the leaf sheaths and leaf blades of the plants. Electrolyte leakage and lipid peroxidation are induced due to the pollutant in the environment which majorly increase the over-production of intracellular reactive oxygen species (Jan *et al.*, 2012) [102]. Triazoles being a group of compound, act both as fungicide as well as plant growth regulator. On one hand it inhibited lipid membranes and electrolyte leakage in carrot plant as found in the study done by Gopi *et al.* (2007) [33] while on the other hand an increase in *Cucurbita maxima* L. seedlings under

paraquat application by foliar spray (50-1000 μM) was found to behave as plant growth regulator as reported by Yoon *et al.* (2011) [104]. 2,4-dichlorophenols and pentachlorophenols are herbicides when applied in soil at 0-5 mg Kg^{-1} in *Triticum aestivum* L. resulted in increased TBARS, free phenols content and guaiacol POD activity but decreased CAT and SOD activity (Michalowicz *et al.*, 2009) [62]. Moreover, in mung bean seedlings when chlorpyrifos was applied on foliar at dosage of 0.3mM showed a significant increase in the yield and its attributing characters viz. highest number of pods, number of seeds per plant and dry seed weight (Parween *et al.*, 2012) [74].

Unselective use of pesticides leads to high residue levels in food. Even a tinch of level present in food becomes very dangerous for human health as it get accumulated in our adipose tissue when taken over a long period of time (Sanborn *et al.*, 2004) [82]. The Maximum Residue Limits of pesticides in food intake as food morals diverge widely for the same pesticide on the same product among countries as well as with the international Codex Committee standards (Codex, 2010) [19]. However, still scientist are struggling hard to get safe level of pesticides residue in the food because many of the cell process inside the bodies work greatly with the help of chemical messengers at very small quantities in ppm or even ppb (Boobis *et al.*, 2008) [12]. Dhas and Srivastava (2010) [20] studied the effect of carbaryl on brinjal fruits where they noticed that 9.93 ppm of carbaryl on 0.2% foliar spray get transferred to 11.47 ppm within a single day after treatment and made a shrinkage in residue to about 13.40 %. Therefore, studies should be undertaken on the side effects of pesticides, its persistence level in crops and its following effects on soil microbial flora, properties and associated nitrogen metabolism.

Nitrogen and Phosphorus

Nitrogen as well as phosphorus is required by the plants in huge amount. According to FAO, global consumption of fertilizers will reach to a huge amount of 200 million tons by 2018. The major countries that cover over 50 % of the fertilizers globally are the China, India and the United States of America (FAO, 2015) [24].

In most of the living structures of the cell like DNA, RNA, hormones, proteins, enzymes and vitamins, nitrogen and phosphorus are an essential component. They are available at many oxidation states both in organic and inorganic form. Through microbial activity unreactive form such as gaseous nitrogen (N_2) is assimilated, ammonium (NH_4^+) and nitrate (NO_3^-) forms are utilized by plants and the complex forms like nucleic acids and amino acids are required by the animals (Yaron *et al.*, 2012) [102]. Phosphorus is used to transport the energy in the form of adenosine triphosphate (ATP) and is also the main macronutrient of living organisms. But when these two components are present in the excess amount in agricultural soils in form of fertilizers become pollutants and cause destruction in the environment (Carpenter *et al.*, 1998 [15]; Torrent *et al.*, 2007) [97]. By leaching and surface run-off these nutrients get into the groundwater and transported to the water bodies which leads to the accumulation of high nitrate content causing eutrophication which ultimately leads to the associated environmental and human health problems (Pretty *et al.*, 2003; Yaron *et al.*, 2012 [102]; Frumin and Gildeeva 2014) [26]. These nitrogen and phosphate fertilizers also have many heavy metals as As, Cd, Cr, Hg, Pb, and Zn which are

harmful for soil as well as plants (Brevik, 2013) and when these nutrients applied in excess amount have negative effect on the yield. As nitrogen is an important constituent of chlorophyll, providing high energy for flower and bud growth, essential for the root elongation and foliage proliferation also makes plant more prone to pathogen attack. Hao *et al.* (2003) [37] reported that it affects the balance of crop nutrients. Nitrogen in high amount increases the decomposition of soil organic matter which directly affects the microbial community activities and its composition (Shen *et al.*, 2010 [85]; Zhou and Zhang, 2014 [106]; Luo *et al.*, 2017) [55] along with the soil salinity and acidity (Han *et al.*, 2015) [36]. Singh and Ghoshal (2010) [89] found that nitrogen when applied in form of organic soil amendments in agricultural soil masks the negative effect of herbicides on soil fertility in terms of soil microbial biomass leading to considerable increase soil fertility and productivity in turn.

Radioactive Contamination

Radioactive contamination refers to the undesirable presence of radioactive elements or compounds in the biota (Smičiklas and Šljivić-Ivanović, 2016). The significant addition of radionuclides to the environment started with the construction and operation of nuclear reactors from 1945 onwards. Apart from the nuclear discharge, the use of fertilizers in heavy amount can deliberately affect the levels of naturally occurring radionuclides, more specifically the use of nitrogen, phosphorus and potassium. Uncontrolled exploitation of the nuclear power acts as a major source of pollution which can enter and affect the environment. The soils which are heavily contaminated with radioactive substances lose their efficiency to produce better quality crops and thus can be classified as degraded (Miura, 2016) [64]. For example the excavation and processing of uranium ore, over production and recycling of the nuclear fuels to the processing and disposal of radioactive wastes leads to heavy degradation of the agricultural land interferes with the crop productivity (Wuana and Okieimen, 2011).

The production of these radioactive wastes primarily occurs by the generation of nuclear power, various industries, medicines, agriculture system and research etc. (Ivana Smičiklas and Marija Šljivić-Ivanović, 2016). These pollutants inhibit the growth of the plant growing in the contaminated soil. Roots act as the primary target site in the translocation of these harmful radionuclides inside the plant and causes severe disturbance in the metabolism (Golmakani *et al.*, 2008 [32]; Singh and Prasad, 2019) [91]. The presence of radioactive substances disturbs PSII photochemistry and enhanced production of reactive oxygen species which ultimately diminishes the overall performance of the plant leading to the decay of the plant with less crop productivity (Hong *et al.*, 2019) [38]. In *Koeleria gracilis* Pers. it was studied that the frequency of aberrant cells and chromosome aberrations in the apical meristem of germinated seeds was significantly increased with respect to plants at other plots with lower levels of radioactive contamination. These results emphasize the role of radioactive contamination in the mutagenic and cytogenetic effects (Geraskin *et al.*, 2013) [28].

Microbial Pollution

Pathogenic bacteria present in the soil makes the soil less inhabitable for the crops growing there (Vukicevich *et al.*, 2016) [99]. Contamination of soil and crops by pathogenic

agents is the effect of waste water reuse in agriculture that receives most attention from environmentalists and scientists. Discharge of effluents from municipal and industrial waste water contains a broad variety of bacteria, protozoa and viruses coming through human and animal feces and urine (Oguis *et al.*, 2019) [70]. Therefore, this water acts as an agent of intestinal infection which either comes through contact or ingestion of soil or indirectly via ingestion of polluted crops (Pandey *et al.*, 2014) [72]. People at high risk include farmers and their families, crop handlers, product consumers and inhabitants of the area nearby to the irrigated fields (Cifuentes *et al.*, 2000 [17], Mara *et al.*, 2007) [60]. Among all these groups most affected ones are agricultural workers since they are more blindly exposed to contaminated soils (WHO, 2006) [101].

Crops being sessile in nature get easily polluted due to their direct contact with waste water during irrigation (Brega *et al.*, 2003) [13]. Contamination of the edible plant parts depend not only on the quality of water, but also on the quantity and the method of its application to soil and also on the type of crop i.e. the ratio of contamination with microbes varies from plants to plants (Jaramillo and Restripo, 2017). The pathogens attack the plants through various pathways (Fig. 2). When zucchini plants were spray- irrigated with waste water then the plants accumulated more pathogens on their surface rather

than other crops. This in part happens due the hairy appearance and a close proximity of the plant to the soil which facilitates better attachment to pathogens (Oguis *et al.*, 2019) [70]. The method of entry, translocation and survival period of the microbes inside the plant also differs (Zhang *et al.*, 2009) [105]. *E. coli* can be translocated from soil to the leaves of lettuce through the root system or through wounds in vegetal tissues (Mandrell *et al.*, 2011) [58]. Wounds also allow the entrance of *Salmonella* and *E. coli* to lettuce and tomato plants (Liu *et al.*, 2018) [54]. The non-pathogenic *E. coli* can survive in soil for one month whereas in spinach leaves the pathogenic strain of *E. coli* O157:H7 can stay for 14 days (Zhang *et al.*, 2009) [105]. It is reported that survival of pathogenic bacteria can increase by internalization within the plant tissues (Lim *et al.*, 2014). Similarly, it is also documented that *E. coli* can use the stomatal cavities in leaves to enter the internal structure of leaves. Inside the plant tissues, rate of survival of pathogens increase considerably since they use cellulose as their main source of carbon except few protozoans since they are larger in size than bacteria thus cannot access the internal parts of the plant; however, they can easily adhere to the surface of edible parts by the excretion of polymers which facilitate adhesion (Ojuederie and Babalola, 2017) [71].

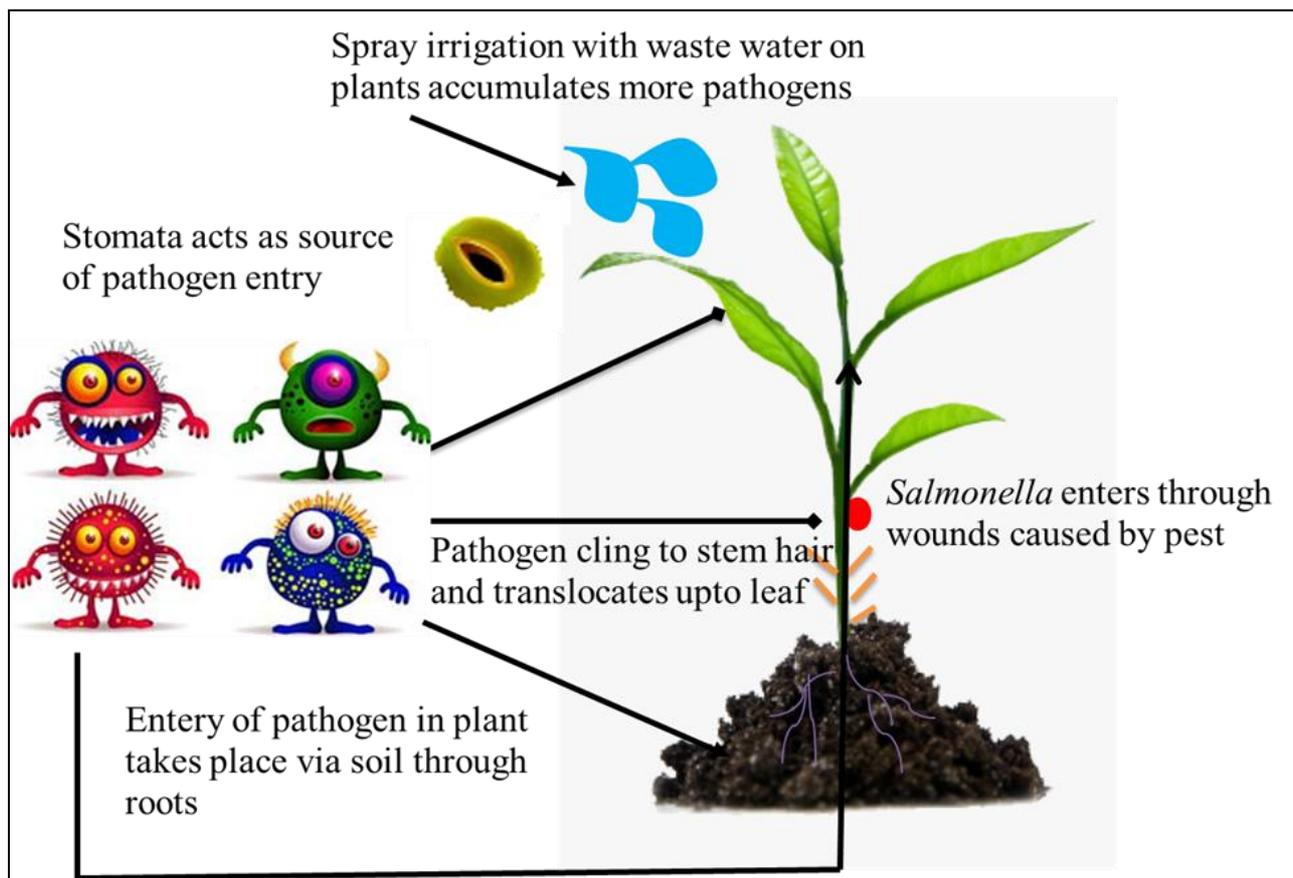


Fig 2: Sources and causes of microbial pollution in plants.

Conclusion

To meet the increasing food demands, agriculture is the best medium to work with and soil being the only medium to practice agriculture. But soil as an important part of cultivated lands is severely affected by agricultural practices. Extensive and non-proportional use of chemical fertilizers and pesticides

has contributed to soil pollution which directly or indirectly affects the plant growth and metabolism. Also, soil ecosystem is hampered by the use of toxic and persistent heavy metals which had tremendously affected the soil physico-chemical and biological properties. Salt is also a major input in interfering the ionic imbalance in the plant system also

damaging the soil by increasing the sodicity and alkalinity which directly hampers the overall growth and productivity of the cash crops. Soil is heavily contaminated by radioactive substances which share weaker bond with soil components that involve higher mobility of pollutant which easily get adsorbed by the biota and accumulates which enhance the level of contamination. Microbial populations constitute major shift in the structure and composition of soil properties and thereby affects functionality of soil and natural food web. Thus, these all above constituents damage the soil quality and fertility and vast biodiversity of the agricultural lands. Hence, unlike these toxic chemical inputs, cost effective and environmental friendly organic amendments must be adopted as a remedial strategy that improves the overall quality and fertility of soil which will be helpful for sustainable agricultural practices.

Acknowledgements

The authors greatly acknowledge the Head, Department of Botany for providing all laboratory facilities. The authors Divya Gupta and Abreeq Fatima are grateful for financial support by the University Grants Commission, Govt. of India in form of Research scholarship. Pratibha Singh is thankful to the Department of Science and Technology, New Delhi, Govt. of India for post doctoral fellowship in form of women scientist (Individual Postdoctoral Grant No. DST/WOS b/SoRF-PM/062/2013/G). Sunita Kumari Singh gratefully acknowledges the Head, Department of Botany, Government College Rajpur, Balrampur, Chattisgarh, India.

References

- Ahmad I, Akhtar M.J, Zahir Z.A, Jamil A. Effect of Cadmium on Seed Germination and Seedling Growth of Four Wheat (*Triticum aestivum* L.) Cultivars *Pakistan Journal of Botany*. 2012; 44:1569-1574.
- Akbarimoghaddam H, Galavi M, Ghanbari A, Panjehkeh N. Salinity Effects on Seed Germination and Seedling Growth of Bread Wheat Cultivars *Trakia Journal of Sciences*. 2011; 9:43-50.
- Alloway B.J. Heavy Metal in Soils. John Wiley and Sons, New York, NY, USA, 1990.
- Ashraf M, Harris P.J.C. Potential Biochemical Indicators of Salinity Tolerance in Plants *Plant Science*. 2004; 166:3-16.
- Assche F, Clijsters H. Effects of Metals on Enzyme Activity in Plants *Plant, Cell and Environment*. 1990; 24:1-15.
- Baath E. Effects of Heavy Metals in Soil on Microbial Processes and Populations (a review) *Water, Air, and Soil Pollution*. 1989; 47:335-379.
- Baker A.J.M. Accumulators and Excluders Strategies in the Response of Plants to Heavy Metals *Journal of Plant Nutrition*. 1981; 3:643-654.
- Bano A, Fatima M. Salt Tolerance in *Zea mays* (L.) Following Inoculation with *Rhizobium* and *Pseudomonas* *Biology and Fertility of Soils*. 2009; 45:405-413.
- Barry P, Young A.J, Briton G. Photodestruction in Higher Plants by Herbicide Action *Journal of Experimental Botany*. 1990; 41:123-126.
- Basantani M, Srivastava A, Sen S. Elevated Antioxidant Response and Induction of Tau-class Glutathione S-Transferase after Glyphosate Treatment in *Vigna radiata* (L.) Wilczek *Pesticide Biochemistry and Physiology*. 2011; 99:111-117.
- Blaylock M.J, Huang J.W. Phytoextraction of Metals. In: I. Raskin and B.D. Ensley (Eds). *Phytoremediation of toxic metals using plants to clean up the environment* Wiley, New York, NY, USA, 2000, 53-70.
- Boobis A.R, Ossendorp B.C, Banasiak U, Hamey P.Y, Sebestyen I, Moretto A. Cumulative Risk Assessment of Pesticide Residues in Food *Toxicology Letters*. 2008; 15:137-150.
- Brega Filho D, Mancuso P.C, Reúso de Água. Universidade de São Paulo-Facultade de Saúde Pública: São Paulo Brasil, 2003, 579.
- Brevik E.C. Soils and Human Health: An overview. In: E.C. Brevik and L.C. Burgess (Eds). *Soils and Human Health* CRC Press, 2013, 29-58.
- Carpenter S.R., Caraco N.F, Correll D.L, Howarth R.W, Sharpley A.N, Smith V.H. Nonpoint Pollution of Surface Water with Phosphorus and Nitrogen *Ecological Applications*. 1998; 8:559-568.
- Castaldi S, Rutigliano F.A, Virzo de Santo A. Suitability of Soil Microbial Parameters as Indicators of Heavy Metal Pollution *Water, Air, and Soil Pollution*. 2004; 158:21-35.
- Cifuentes E, Gomez M, Blumenthal U, Tellez-Rojo M.M, Romieu I, Ruiz-Palacios G, Ruiz-Velazco S. Risk Factors for *Giardia Intestinalis* Infection in Agricultural Villages Practicing Wastewater Irrigation in Mexico *American Journal of Tropical Medicine and Hygiene*. 2000; 62:388-392.
- Clarkson D.T, Luttge U. Mineral Nutrition: Divalent Cations, Transport and Compartmentation *Progress in Botany*. 1989; 51:93-112.
- Codex. Codex aliment Arius commission pesticide residues in food and feed Available from, 2010. <http://www.codexalimentarius.net/pestres/data/pesticides/index.html>.
- Dhas S, Srivastava M. An Assessment of Carbaryl Residues on Brinjal Crop in an Agricultural Field in Bikaner, Rajasthan (India) *Asian Journal of Agricultural Sciences*. 2010; 2:15-17.
- Djingova R, Kuleff I. Instrumental Techniques for Trace Analysis. In: J.P Vernet (Ed). *Trace Elements their distribution and effects in the environment* London, UK, Elsevier, 2000.
- Dubey K.K, Fulekar M.H. Effect of Pesticides on the Seed Germination of *Cenchrus setigerus* and *Pennisetumpedicellatum* as Monocropping and Co-cropping System: Implications for Rhizospheric Bioremediation *Romanian Biotechnological Letters*. 2011; 16:5909-5918.
- FAO. Food Security and Agricultural Mitigation in Developing Countries: Options for Capturing Synergies Rome, Italy, Retrieved January 26, 2019, from, 2009. <http://www.fao.org/docrep/012/i1318e/i1318e00.pdf>.
- FAO. World fertilizer trends and outlook to 2018. Rome, Food and Agriculture Organization of the United Nations Statistics. Retrieved January 26, 2019, from, 2015. <http://www.fao.org/3/a-i4324e.pdf>.
- Friedlová M. The Influence of Heavy Metals on Soil Biological and Chemical Properties *Soil and Water Research*. 2010; 5:21-27.
- Frumin G.T, Gildeeva I.M. Eutrophication of Water Bodies - A Global Environmental Problem *Russian*

- Journal of General Chemistry*. 2014; 84:2483-2488.
27. Fuerst E.P, Norman M.A. Interactions of Herbicides with Photosynthetic Electron Transport *Weed Science*. 1991; 39:458-464.
 28. Geraskin S, Evseeva T, Oudalova A. Effects of Long-term Chronic Exposure to Radionuclides in Plant Populations *Journal of Environmental Radioactivity*. 2013; 121:22-32.
 29. Gang Li, Nan Lu, Yang W, Zhu D. Relationship between Heavy Metal Content in Polluted Soil and Soil Organic Matter and pH in Mining Areas *Materials Science and Engineering*. 2018; 394(5):052081.
 30. Ghani A. Toxic Effects of Heavy Metals on Plant Growth and Metal Accumulation in Maize (*Zea mays* L.) *Iranian Journal of Toxicology*. 2010; 3:325-334.
 31. Giller K.E, Witter E, Mcgrath S.P. Toxicity of Heavy Metals to Microorganisms and Microbial Processes in Agricultural Soils *Soil Biology and Biochemistry*. 1998; 30:1389-1414.
 32. Golmakani S, Moghaddam M.V, Hosseini T. (Factors Affecting the Transfer of Radionuclides from the Environment to Plants *Radiation Protection Dosimetry*. 2008; 130(3):368-375.
 33. Gopi R, Jaleel C.A, Sairam R, Lakshmanan Gomathinayagam M, Panneerselvam. Differential Effects of Hexaconazole and Paclobutrazol on Biomass, Electrolyte Leakage, Lipid Peroxidation and Antioxidant Potential of *Daucus carota* L *Colloids and Surfaces B Biointerfaces*. 2007; 60:180-186.
 34. Guignard M.S, Leitch A.R, Acquisti C, Eizaguirre C, Elser J.J, Hessen D.O *et al*. Impacts of Nitrogen and Phosphorus: From Genomes to Natural Ecosystems and Agriculture *Frontiers in Ecology and Evolution* 5 article 2017; 70:1-9.
 35. Haddad S.A, Lemanowicz J, Abd El-Azeim M.M. Cellulose Decomposition in Clay and Soils Contaminated with Heavy Metals *International Journal of Environmental Science and Technology*. 2018; 16(7):3275-3290.
 36. Han J, Shi J, Zeng L, Xu J, Wu L. Effects of Nitrogen Fertilization on the acidity and Salinity of Greenhouse Soils *environmental science and pollution research*. 2015; 22:2976-2986.
 37. Hao X, Chang C, Travis G.R, Zhang F. Soil Carbon and Nitrogen Response to 25 Annual Cattle Manure Applications *Journal of Plant Nutrition and Soil Science*. 2003; 166:239-245.
 38. Hong D.D, Anh H.T.L, Tam L.T, Show P.L, Hui Y.L. Effects of Nanoscale Zerovalent Cobalt on Growth and Photosynthetic Parameters of Soybean *Glycine max* (L.) MerrDT26 at Different Stages *BMC Energy* 1 article 6, 2019.
 39. Hu Y, Schmidhalter U. Limitation of Salt Stress to Plant Growth In B Hock and C.F Elstner (Eds), *Plant Toxicology* New York, Marcel Dekker Inc, 2002, 91-224.
 40. Isayenkov S.V, Maathuis F. Plant Salinity Stress; Many Unanswered Questions Remain *Frontiers in Plant Science*. 10 article 80, 2018.
 41. Jacoby R, Peukert M, Succurro A, Koprivova A, Kopriva S. The Role of Soil Microorganisms in Plant Mineral Nutrition - Current Knowledge and Future Directions, *Frontiers in Plant Science* 8 article 1617, 2017, 1-19.
 42. Jamil A, Riaz S, Ashraf M, Foolad M.R. Gene Expression Profiling of Plants under Salt Stress, *Critical Reviews in Plant Sciences*. 2011; 30:435-458.
 43. Jan S, Parween T, Siddiqi T.O, Mahmooduzzafar. Antioxidant Modulation in Response to Gamma Induced Oxidative Stress in Developing Seedlings of *Psoraleacorylifolia* L, *Journal of Environmental Radioactivity*. 2012; 113:142-149.
 44. Jaramillo M.F, Restrepo I. Wastewater Reuse in Agriculture: A Review about its Limitations and Benefits. *Sustainability*. 2007; 9:1734-1740.
 45. Javid M.G, Sorooshzadeh A, Moradi F, SanavySeyed A.M.M, Allahdadi I. The Role of Phytohormones in Alleviating Salt Stress in Crop Plants, *Australian Journal of Crop Science*. 2011; 5:726-734.
 46. Jayakumar K, Jaleel C.A, Azooz M.M. Phytochemical Changes in Green Gram (*Vigna radiata*) under Cobalt Stress, *Global Journal of Molecular Sciences*. 2008; 3:46-49.
 47. Jayakumar K, Jaleel C.A, Vijayarengan P. Changes in Growth, Biochemical Constituents, and Antioxidant Potentials in Radish (*Raphanus sativus* L) under Cobalt Stress *Turkish Journal of Biology*. 2007; 31:127-136.
 48. Jayakumar K, Rajesh M, Baskaran L, Vijayarengan P. Changes in Nutritional Metabolism of Tomato (*Lycopersicon esculantam* Mill.) Plants Exposed to Increasing concentration of cobalt Chloride, *International Journal of Food Nutrition and Safety*. 2013; 4:62-69.
 49. Joy M, Abit M, Al Khatib K. Metabolism of Quizalofop and Rimsulfuron in Herbicide Resistant Grain Sorghum, *Pesticide Biochemistry and Physiology*. 2013; 105:24-27.
 50. Kabata-Pendias A. Trace Elements in Soils and Plants, CRC Press, Boca Raton, Fla, USA, 3rd edition, 2001.
 51. Khan H, Zeb A, Ali Z, Shah S.M. Impact of Five Insecticides on Chickpea (*Cicer arietinum*L.) Nodulation, Yield and Nitrogen Fixing Rhizospheric Bacteria *Soil and Environment*. 2009; 28:56-59.
 52. Kibra M.G. Effects of Mercury on Some Growth Parameters of Rice (*Oryza sativa* L) *Soil and Environment*. 2008; 27:23-28.
 53. Lim J.A, Lee D.W, Heu S. The Interaction of Human Enteric Pathogens with Plants *Journal of Plant Pathology*. 2014; 30:109-116.
 54. Liu H, Whitehouse C.A, Li B. Presence and Persistence of *Salmonella* in Water: the Impact on Microbial Quality of Water and Food Safety *Frontiers in Public Health*. 2018; 6:159.
 55. Luo L, Meng H, Wu R, Gu J.D. Impact of Nitrogen Pollution/ Deposition on Extracellular Enzyme Activity, Microbial Abundance and Carbon Storage in Coastal Mangrove Sediment *Chemosphere*. 2017; 177:275-283.
 56. Luo Y, Rimmer D.L. Zinc-Copper Interaction Affecting Plant Growth on a Metal-Contaminated Soil *Environmental Pollution*. 1995; 88:79-83.
 57. Mahmood K.S, Shah Z.A. Screening of the Best Insecticide in Reducing the Chickpea Pod Borer Damage Infected by Gram Pod Borer (*H. armigera*) in Faisalabad *Pakistan Journal of Biological Sciences*. 2003; 6:1156-1158.
 58. Mandrell R.E. Tracing pathogens in fruit and vegetable production chains In: S Brul P.M Fratamico T McMeekin (Eds), *Tracing pathogens in the food chain* Cambridge, UK, Woodhead Publishing Ltd, 2011, 548-595.
 59. Manivasagaperumal R, Balamurugan S, Thiyagarajan G, Sekar J. Effect of zinc on germination, seedling growth

- and Biochemical Content of Cluster Bean (*Cyamopsis tetragonoloba* (L) Taub *Current Botany*. 2011; 2:11-15.
60. Mara D.D, Sleigh P.A, Blumenthal U.J, Carr R.M. Health Risks in Wastewater Irrigation: Comparing Estimates from Quantitative Microbial Risk Analyses and Epidemiological Studies *Journal of Water and Health*. 2007; 5:39-50.
 61. Marques A.P.G.C, Rangel A.O.S.S, Castro P.M.L. Remediation of Heavy Metal Contaminated Soils: Phytoremediation as a Potentially Promising Clean-up Technology *Critical Reviews in Environmental Science and Technology*. 2009; 39:622-654.
 62. Michalowicz J, Posmyk M, Duda W. Chlorophenols Induce Lipid Peroxidation and Change Antioxidant Parameters in the Leaves of Wheat (*Triticum aestivum* L), *Journal of Plant Physiology*. 2009; 166:559-568.
 63. Mishra V., Srivastava G, Prasad S.M, Abraham G. Growth, Photosynthetic Pigments and Photosynthetic Activity during Seedling Stage of Cowpea (*Vigna unguiculata*) in Response to UV-B and Dimethoate, *Pesticide Biochemistry and Physiology*. 2008; 92:30-37.
 64. Miura S. The Effects of Radioactive Contamination on the Forestry Industry and Commercial Mushroom-log Production in Fukushima, Japan. In Nakanishi T, Tanoi K. (Eds) *Agricultural implications of the Fukushima Nuclear Accident Tokyo*, Springer, 2016, 145-160.
 65. Moore M.T, Kroger R. Effect of Three Insecticides and Two Herbicides on Rice (*Oryza sativa*) Seedling Germination and Growth, *Archives of Environmental Contamination and Toxicology*. 2010; 59:574-581.
 66. Munns R. Genes and Salt Tolerance: Bringing them together *New Phytologist*. 2005; 167:645-663.
 67. Munns R, Rawson H.M. Effect of Salinity on Salt Accumulation and Reproductive Development in the Apical Meristem of Wheat and Barley, *Australian Journal of Plant Physiology*. 1999; 26:459-464.
 68. Netondo G.W, Onyango J.C, Beck E. Sorghum and Salinity: II. Gas Exchange and Chlorophyll Fluorescence of Sorghum under Salt Stress *Crop Sciences*. 2004; 44:806-811.
 69. Nicholls A.M, Mal T.K. Effects of Lead and Copper Exposure on Growth of an Invasive Weed, *lythrum salicaria* l. (purple loosestrife) *Ohio Journal of Science*. 2003; 103:129-133.
 70. Oguis G.K, Gilding E.K, Jackson M.A, Craik D.J. Butterfly Pea (*Clitoria ternatea*), A Cyclotide-Bearing Plant with Applications in Agriculture and Medicine *Frontiers in Plant Science* 10 article 645, 2019, 1-23.
 71. Ojuederie O.B, Babalola O.O. Microbial and Plant-Assisted Bioremediation of Heavy Metal Polluted Environments: A Review, *International Journal of Environmental Research and Public Health*. 2017; 14:1504.
 72. Pandey P.K, Kass P.H, Soupir M.L, Biswas S, Singh V.P. Contamination of Water Resources by Pathogenic Bacteria *AMB Express*. 2014; 4:51.
 73. Parihar P, Singh S, Singh R, Singh V.P, Prasad S.M. Effect of Salinity Stress on Plants and its Tolerance Strategies: A Review *Environmental Science and Pollution Research*. 2015; 22:4056-4075.
 74. Parween T, Jan S, Mahmooduzzafar, Fatma T. Evaluation of Oxidative Stress in *Vigna radiata* L in Response to Chlorpyrifos, *International Journal of Environmental Science and Technology*. 2012; 9:605-612.
 75. Parween T, Jan S, Mahmooduzzafar, Fatma T. Assessing the Impact of Chlorpyrifos on Growth, Photosynthetic Pigments and Yield in *Vigna radiata* L at Different Phenological Stages, *African Journal of Agricultural Research*. 2011; 6:4432-4440.
 76. Patel B.B, Patel Bharat B, Dave R.S. Studies on Infiltration of Saline - Alkali Soils of Several Parts of Mehsana and Patan Districts of North Gujarat, *Journal of Applied Technology and Environmental Sanitation*. 2011; 1:87-92.
 77. Pretty J.N, Mason C.F, Nedwell D.B, Hine R.E, Leaf S, Dils R. Environmental Costs of Freshwater Eutrophication in England and Wales, *Environmental Science and Technology*. 2003; 37:201-208.
 78. Rahneshan Z, Nasibi F, Moghadam A.A. Effects of Salinity stress on Some Growth, Physiological, Biochemical Parameters and Nutrients in two Pistachio (*Pistacia vera* L.) Rootstocks, *Journal of Plant Interactions*. 2018; 13:73-82.
 79. Rajashekar N, Prakasha, Murthy T.C.S. Seed Germination and Physiological Behavior of Maize (Cv. NAC-6002) Seedlings under Abiotic Stress (Pendimethalin) Condition, *Asian Journal of Crop Science*. 2012; 4:80-85.
 80. Raskin I, Nanda-Kumar P.B.A, Dushenkov S, Salt D.E. Bioconcentration of Heavy Metals by Plants, *Current Opinion in Biotechnology*. 1994; 5:285-290.
 81. Salgare S.A, Acharekar C. Effect of Industrial Pollution on Growth and Content of Certain Weeds, *Journal of Nature Conservation*. 1992; 4:1-6.
 82. Sanborn M, Cole D, Kerr K, Vakil C, Sanin L.H, Basil K. Systematic Review of Pesticides Human Health Effects. The ontario, college of family physicians. Retrieved December 27, 2018, from, 2004. <http://www.ocfp.on.ca/local/files/Communications/Current%20Issues>.
 83. Shahbaz M, Ashraf M. Improving Salinity Tolerance in Cereals, *Critical Reviews in Plant Sciences*. 2013; 32:237-249.
 84. Sharma M.S.R, Raju N.S. Correlation of Heavy Metal Contamination with Soil Properties of Industrial areas of Mysore, Karnataka, India by Cluster Analysis. *International Research Journal of Environmental Sciences*. 2013; 10:22-27.
 85. Shen W, Lin X, Shi W, Min J, Gao N, Zhang H *et al*. Higher Rates of Nitrogen Fertilization Decrease Soil Enzyme Activities, Microbial Functional Diversity and Nitrification Capacity in a Chinese Poly tunnel Greenhouse Vegetable land *Plant Soil*. 2010; 337:137-150.
 86. Shen Z, Li X, Wang C, Chen H, Chua H. Lead Phytoextraction from Contaminated Soil with High-Biomass Plant Species, *Journal of Environmental Quality*. 2002; 31:1893-1900.
 87. Silva V, Mol H.G.J, Zomer P, Tienstra M, Ritsema C.J, Geissen V. Pesticide Residues in European Agricultural Soils - A Hidden Reality Unfolded, *Science of the Total Environment*. 2019; 653:1532-1545.
 88. Singh M, Singh V.P, Prasad S.M. Nitrogen Alleviates Salinity Toxicity in *Solanum lycopersicum* Seedlings by Regulating ROS Homeostasis, *Plant Physiology and Biochemistry*. 2019; 141:166-176.

89. Singh P, Ghoshal N. Variation in Total Biological Productivity and Soil Microbial Biomass in Rainfed Agro ecosystems: Impact of Application of Herbicide and Soil Amendments, *Agriculture Ecosystem & Environment*. 2010; 137:241-250.
90. Singh P, Prasad S.M. Antioxidant Enzyme Responses to the Oxidative Stress due to Chlorpyrifos, Dimethoate and Dieldrin Stress in Palak (*Spinacia oleracea* L.) and their Toxicity Alleviation by Soil Amendments in Tropical Croplands *Science of the Total Environment*. 2018; 630:839-848.
91. Singh S, Prasad S.M. Management of Chromium (VI) Toxicity by Calcium and Sulfur in Tomato and Brinjal: Implication of Nitric Oxide, *Journal of Hazardous Material*. 2019; 373:212-223.
92. Smejkalova M, Mikanova O, Boruvka L. Effects of Heavy Metal Concentrations on Biological Activity of Soils Microorganisms *Plant, Soil and Environment*. 2003; 49:321-326.
93. Smičiklas I, Šljivić-Ivanović M. Radioactive Contamination of the Soil Assessments of Pollutants Mobility with Implication to Remediation Strategies, In: M.L Larramendy, S. Soloneski (Eds), *Soil contamination - Current consequences and further solutions Intech Open* DOI: 10.5772/64735, 2016.
94. Suri K.S, Singh G. Insecticide Induced Resurgence of the White Backed Plant Hopper *Sogatella furcifera* (Horvath) (Hemiptera: Delphacidae) on Rice Varieties with Different Levels of Resistance *Crop Protection*. 2011; 30:118-124.
95. Tabur S, Demir K. Role of Some Growth Regulators on Cytogenetic Activity of Barley under Salt Stress, *Plant Growth Regulation*. 2010; 60:99-104.
96. Taiz L, Zeiger E. *Plant Physiology* Sinauer Associates, Sunderland, Mass, USA, 2002.
97. Torrent J, Barberis E, Gil-Sotres F. Agriculture as a Source of Phosphorus for Eutrophication in Southern Europe *Soil Use Management*. 2007; 23:25-35.
98. Tort N, Turkyilmaz B. Physiological Effects of Captan Fungicide on Pepper (*Capsicum annuum* L.), *Pakistan Journal of Biological Sciences*. 2003; 6:2026-2029.
99. Vukicevich E, Lowery T, Bowen P. Cover Crops to Increase Soil Microbial Diversity and Mitigate Decline in Perennial Agriculture - A Review *Agronomy for Sustainable Development*. 2016; 36(3):48.
100. Wang A.S, Angle J.S, Chaney R.L, Delorme T.A, Reeves R.D. Soil pH Effects on Uptake of Cd and Zn by *Thlaspi caerulescens* Plant Soil. 2006; 281:325-337.
101. WHO. Guidelines for the Safe Use of Wastewater, Excreta and Greywater Geneva, 2006.
102. Yaron B, Dror I, Berkowitz B. Soil-Subsurface Change Chemical pollutant impacts berlin, Heidelberg, springer berlin Heidelberg, Retrieved on January 25, 2019, from, 2012.
<http://link.springer.com/10.1007/978-3-642-24387-5>.
103. Yi L, Hong Y, Wang D, Zhu Y. Determination of Free Heavy Metal Ion Concentrations in Soils Around a Cadmium Rich Zinc Deposit *Geochemistry Journal*. 2007; 41:235-240.
104. Yoon J.Y, Shin J.S, Shin D.Y, Hyun K.H, Burgos N.R, Sungbeom L. Tolerance to Paraquat-Mediated Oxidative and Environmental Stresses in Squash (*Cucurbita* spp.) Leaves of various Ages *Pesticide Biochemistry and Physiology*. 2011; 99:65-76.
105. Zhang G, Ma L., Beuchat L.R, Erickson M. Lack of Internalization of *Escherichia coli* O157:H7 in Lettuce (*Lactuca sativa* L.) after Leaf Surface and Soil Inoculation, *Journal of Food Protection*. 2009; 72:2028-2037.
106. Zhou X, Zhang Y. Temporal Dynamics of Soil Oxidative Enzyme Activity Across a Simulated Gradient of Nitrogen Deposition in the Gurbantunggut Desert, Northwestern China *Geoderma*. 2014; 213:261-267.
107. Zhu J.K. Salt and Drought Stress Signal Transduction in Plants *Annual Review of Plant Biology*. 2002; 53:247-273.