



ISSN 2347-2677

www.faunajournal.com

IJFBS 2020; 7(4): 06-08

Received: 06-05-2020

Accepted: 09-06-2020

Ashiya

Research Scholar, Department of
Zoology, J.P. University,
Chapra, Bihar, India

Biochemical changes in Air-breathing fish, *Anabas testudineus* (Bloch) exposed to the Pesticide Rogor (Dimethoate)

Ashiya

Abstract

In this paper the study has been designed to determine the LC₅₀ values of Rogor an insecticide and relative behavioral changes in fresh water fish *Anabas testudineus* (Bloch) exposed for 24, 48, 72 and 96 hrs. The LC₅₀ values for *Anabas testudineus* were found at 9ppm, 8.31ppm, 7.8ppm and 7.1ppm respectively. Fish show increased opercular moment, loss of equilibrium, increased surface activity, over secretion of mucous, irregular swimming activity, rapid jerky movements and aggressiveness were observed.

Keywords: Biochemical changes, Bloch, *Anabas testudineus*, Dimethoate

Introduction

The pesticide application has greatly contributed to meet with the growing demand of food through enhancement of agricultural yields. Excessive use of broad spectrum or non-selective pesticides damages the ecosystem, sometimes irreversibly, contaminate soil surface and groundwater as well as food chains and thus compromises the health and well-beings of aquatic and surrounding terrestrial environments (Bhat *et al.*, 2012; Rita *et al.*, 2006; Veeraiah, 2012) [2, 12, 15]. The toxic discharge from industries, different mining, and agricultural developments and processing find their way to the aquatic environment and may have detrimental effects on the inhabitants (Hamilton and Mehrle, 1986). Pesticides are one of the dangerous pollutants which cause great harm to the animals present in the aquatic environment including fishes.

The toxicity of a pesticide is altered in commercial form and the fish susceptibility also frequently influenced by environmental factors. The knowledge about lethal concentration of any pesticide it is easier to protect its discharge above the limit. Also, toxicity of pesticide may be classified as extreme, high, moderate, slight and relatively harmless chemicals.

Organophosphate compounds comprise insecticides currently used worldwide for agricultural and household applications. These insecticides produce toxicity by inhibition of the enzyme Acetyl cholinesterase accumulates in the central and peripheral nervous system, which, in turn results into over-activation on post-synaptic cholinergic receptors and produce neurotoxicity (Barat and Bahadur, 2014; Muthukumarawel *et al.*, 2013; Pandey *et al.*, 2014) [3, 10]. The organophosphates are more frequently used due to their high insecticidal property, low mammalian toxicity, less persistence and low biodegradation in the environment. Dimethoate is an organophosphate with effect through contact or systemic circulation. The fish serves as a bio-indicator as it responds with great sensitivity to changes in the aquatic environment and thus also had important role in the monitoring of water pollution (Ahmad, 2012; Binukumari and Basanthi, 2013) [1, 4]. The recent and notable work published on the effects of organophosphate on various aspects of fish are those of Das and Mukherjee (2003) [5]; Shrutis and Tantarapale (2016); Tripathi and Rajesh (2015) [14]; Rajani and Revathy (2015) [11].

This investigation will be carried to evaluate the acute toxicity of an organophosphate pesticide-Dimethoate for the carp fish, *Anabas testudineus* due to their easy availability in local waterbodies and their long survival in an aquarium. This is a common edible fish inhabited several type of reservoirs. The insecticide Dimethoate will be selected for study due to its wider application in crop fields. This pesticide is also highly soluble in water and can leach into nearby water sources to affect aquatic organisms. This is a low persistence pesticide with half-life of 4-16 days but lasting also variable with medium conditions.

Corresponding Author:

Ashiya

Research Scholar, Department of
Zoology, J.P. University,
Chapra, Bihar, India

The test fish will be procured and acclimatized in the laboratory. The water quality parameters will evaluate as methods of APHA (2012), and, lethal toxicity for different intervals as standard methods described in monographs.

The biochemical parameters like blood glucose, plasma protein, Serum cholestral, Glutamate Pyruvate Transaminase and Glutamate Oxaloacetate Transaminase will be evaluated using assay kits supplied by scientific emporium, Gorakhpur. The histo-pathological studies for estimation of Glycogen, Protein, Lipid and Cholestral (Pearse, 1995; Bancroft *et al.*, 1994) will be performed in liver, kidney, testes and ovary. The estimation of Alkaline and Acid phosphatase, Pyruvic acid and lactic acid will also be performed (Bancroft *et al.*, 1994).

The one way analysis of variance (ANOVA) was applied to test the significance of difference among the control and experimental values. The P values which were less than 0.5 considered statistically significant.

Materials and Methods

The live freshwater fishes *Anabas testudineus* were collected from river, 35 km away from Siwan district. India and brought to the laboratory. The fishes were fed with live pieces of Earthworm every alternate day and allow acclimatizing in the laboratory conditions in large aquaria for 15 days prior to

the experimentation. Water was renewed every day to provide fresh water rich in oxygen.

The fishes were (average weight 10 gm and length 9.5 cm.) selected for LC₅₀ determination. Preliminary experiment at different concentration of Rogor was conducted to find concentration (ppm) that resulted in 50% mortality in given time. The LC₅₀ values were calculated according to the physico-chemical characteristics of test water have been analyzed during experimentation.

Results and Discussion

The results revealed that the water used for experiment did not contain any toxic substance. Initially no mortality was observed in control group. Fishes exposed to lethal concentrations of Rogor for a short-term exposure were studied in terms of behavioural, rate of survival and mortality. The LC₅₀ values of freshwater fish, *Anabas testudineus* exposed to Rogor for 24, 48, 72 and 96hrs have been recorded at 9 ppm, 8.31ppm, 7.8 ppm and 7.1ppm respectively. The LC₅₀ values, regression results have calculated to support present observations in Table 1. In the present investigation LC₅₀ values were decreased at 96 hrs and found to be increased during the exposure period of 24 hrs. So the values were found at the highest towards 24 hrs and the lowest at 96 hrs of exposure.

Table 1: LC₅₀ values for freshwater *Anabas testudineus* stigma after exposure to insecticide Rogor for a period of 24, 48, 72 and 96 hrs

Exposure Period in hrs.	Regression equation	LC ₅₀ values in % concentrations	Calculated LC 50 values in % concentrations	Variance	Chi-square X'	Fiducial limit up to 95 % confidence
						MIM2
24hrs	Y=11.2266373 X - 5.00094227	9	9	0.00026081	0.00572734	0.92232061 0.98562741
48hrs	Y=7.95149167 X - 5.00225297	8.31	8.3	0.00025651	0.14220305	0.87497733 0.96492473
72hrs	Y=9.24280544 X - 5.00811372	7.8	7.9	0.000433447	0.03248002	0.85734784 0.93896244
96hrs	Y=12.0156952 X - 5.01063729	7.1	7.2	0.00023104	0.11853853	0.82769215 0.88727624

The exposure of fish to different concentrations of rogor shows altered behavioral responses, the restlessness and hyperactivity in fish may occur due to the inactivation of acetylcholinesterase, leading to accumulation of acetylcholine at synaptic junctions. Stimulation of peripheral nervous system which results into increased metabolic activities. Disruption of schooling behaviour of the fish, due to the lethal and sub lethal stress at the toxicant., results in increased swimming activity and entails increased expenditure of energy. The abrupt erratic and jerky swimming may be due to inhibition of the acetylcholinesterase in the brain and neuromuscular junctions.

The opercular movement in fish has been reported to increase following the exposure of toxins. Contrary to it., the opercular movement in fish showed a marked decrease on dimethoate.

Conclusion

In this Study Fishes exposed to lethal concentration of Rogor at 9 ppm for 24 hrs, at first fishes showed increased opercular movement. This could be due to clearance of the accumulated mucus debris in the gill region for proper breathing. At the inception the fishes suffocate and used to come at the surface for gasping the air. The fishes were avoiding toxic water with fast swimming and jumping faster. Presence of over secretion of mucous on the body was found and irregular swimming

activity was noticed.

In this Study changes are the obvious of the motivational biochemical, physiological and environmental influence state of the fishes. An erratic swimming of the treated fishes showed loss of equilibrium. It is prove that the region in the brain associated with the maintenance of equilibrium should have been affected. The erratic swimming, rapid jerky movements and convulsions before death were evident and the serve asphyxiation as indicated by gasping to death. This has been laid to the conclusion that from the present study fishes were highly sensitive to the insecticide Rogor.

References

- Ahmad Z. Toxicity bioassay and effects of sub-lethal exposure of Malathion on biochemical composition and hematological parameters of *Clariens gariepinus*. African Journal of Biotechnology. 2012; 11:8578-8585.
- Bhat BA, Vishwakarma, S Verma, A Saxena G. A comparative study on the toxicity of a synthetic pesticide, dichlorvos and a neem based pesticide, neem-on to *Labeo rohita* (Hamilton). Current World Environment. 2012; 7(1):157-161.
- Barot J, Bahadur A. Toxic effect of azo dye (CI direct green 6) on blood parameters of freshwater fish *Labeo Rohita* (Ham.). Journal of Cell and Tissue Research.

- 2014; 14(2):4251.
4. Binukumari S, Vasanthi J. Studies on the effect of pesticide Dimethoate 30% EC on the lipid content of the fresh water fish *Labeo rohita*; Middle-East Journal of Scientific Research. 2013; 13(2):133-136.
 5. Das BK, Mukherjee SC. Toxicity of cypermethrin in *Labeo rohita* fingerlings: biochemical, enzymatic and haematological consequences. Toxicology and Pharmacology. 2003; 134(1):109-121.
 6. Jaya Shahi, Ajay Singh. Toxic effect of commonly used Synthetic Pesticide on freshwater fish *Clarias batracus*. Trends in Fisheries Research. 2015; 4(2):14-17.
 7. Muthukumaravel K, Rajarajan P, Nathiya N, Govindarajan M, Raveendran S. Studies on the histopathology of selected organ of fresh water fish *Labeo rohita* exposed to pesticide monocrotophos. Biochem Cell Archive. 2013; 10(2):201-206.
 8. Nagaraju B, Venkata Rathnamma V. Affect of profenofos an organophosphate on protein level in some tissues of freshwater fish *Labeo rohita* (Hamilton). Int J Pharm Sci. 2013; 5(Suppl 1):276-290.
 9. Nagpure NS, Kumar R, Kushwaha B, Singh PJ, Srivastava SK. Toxicity Assessment in Fishes: A Practical Approach. National Bureau of Fish Genetic Resource, Lucknow, 2007, 231-257.
 10. Pandey AK, Mishra DK, Bohidar K. Histopathological changes in gonadotrophs of *Channa punctatus* (Bloch) exposed to sublethal concentration of carbaryl and cartap. J Exp. Zool. India. 2014; 17:451-455.
 11. Rajni A, Revathy K. Effect of combined pesticide on acetylcholine esterase activity in freshwater fish *Danio Rerio*: International Journal of Pharma and Biosciences. 2015; 6 (1):1305-1310.
 12. Rita JJ, Arockia NC and Milton MC. Effect of Methonyl on the Biochemical components of the Freshwater *Oreochromis mosambicus*. Ind J Environ & Eco-planning. 2006; 12(1):1-8.
 13. Shruti Gijare, Tantarapale VT. Effects of Cypermethrin on lipid and cholesterol contents of freshwater fish *Channa orientalis* (Bloch). 2014; 3(8):200-209.
 14. Tripathi VK, Rajesh KY. Effect of Pesticide (Organophosphate) on aquatic fish *Labeo Rohita*, International Journal chemical science. 2015; 13(2):626-640.
 15. Veeraiah. Effect of pesticides on non-target organisms. Residue Rev. 2012; 76:173-301.
 16. Visvanathan P, Maruthanayagam C, Govindaraju M. Effect of malathion and endosulfan on biochemical changes in *Channa punctatus*. J Ecotoxicol Environ Monit. 2009; 19:251-7.