



E-ISSN 2347-2677

P-ISSN 2394-0522

<http://www.faunajournal.com>

IJFBS 2023; 10(2): 48-51

Received: 11-02-2023

Accepted: 19-03-2023

TS Pathan

Department of Zoology,
Kalikadevi Arts, Commerce and
Science College, Shirur Kasar,
Beed, Maharashtra, India

SV Rankhamb

Department of Zoology, Late
Ramesh Warpudkar ACS College
Sonpeth, Parbhani,
Maharashtra, India

Shaikh Yasmeen

Department of Zoology, Dr.
Rafiq Zakaria College for
Women, Aurangabad,
Maharashtra, India

Corresponding Author:

Shaikh Yasmeen

Department of Zoology, Dr.
Rafiq Zakaria College for
Women, Aurangabad,
Maharashtra, India

Toxicity and bioaccumulation of cadmium in the freshwater bivalve *Lamellidens marginalis*

TS Pathan, SV Rankhamb and Shaikh Yasmeen

Abstract

The influence of different concentrations of cadmium on Bivalve *Lamellidens marginalis* was examined by a toxicity experiment. Acute toxicity was analyzed by measurement of the 96hrs LC50 and daily survival rates. Results indicated that the mortality rate of *Lamellidens marginalis* was proportional to the concentration of cadmium. LC50 after 96 hrs. the bioaccumulation value of the pollutant in the *Lamellidens marginalis*'s soft parts. The bioaccumulation of cadmium in the organism increased by increasing the concentration of cadmium in solution. The increase of cadmium in the organism was not steady, but it took the form of sudden increase after increasing the concentration of cadmium in solution.

Keywords: *Lamellidens marginalis*, cadmium, bioaccumulation, survival, soft tissue

Introduction

Heavy metal contamination is a serious problem around the world because of the metals' non-biodegradable properties. Their bioavailability and adverse effects on living organisms with long-lasting toxic effects can have environmental risks (Ukoha *et al.* 2014; Sthanadar *et al.* 2015) [40, 39]. They persist indefinitely and cause pollution of the air, water, and soils (Yasmeen, 2019; Shinde, 2021; Yasmeen, *et al.*, 2021) [48, 33, 49]. Thus, the main strategies of pollution control are to reduce the bioavailability, mobility, and toxicity of metals. Heavy metals are natural constituents of the environment, where they are generally found in low concentrations; however, anthropogenic activities contribute to increase their concentration. The impact of these metals in the environment is a serious and growing problem worldwide. Marine pollution has been documented of various kinds of pollutants, especially heavy metals (Romeo *et al.* 2005) [28]. However, some heavy metals are essential for the lives of organisms, while many are highly toxic, and play no significant biological roles (Cross and Sunda 1985; Rainbow 1985; Rainbow and White 1989; Sanders 1997) [12, 25, 51, 30].

Cadmium is a nonessential trace metal that is toxic at low concentrations for aquatic organisms (Cicik and Engin 2005) [10]. Pollution an undesirable changes in the physical, chemical and biological characteristic of air, water and soil, which is harmful to living organisms. Whatever may be the mode of contamination polluted environment is what suitable for existing life forms.

Cadmium is a metallic contaminant that has no known essential functions of human physiology (Barak & Mason, 1990) [5]. It has gained wide interest in the scientific community in recent years due to its potential human health hazards (Selvi *et al.*, 2003) [31]. The main source of cadmium pollution is through soil. Cadmium released from factory wastes and mining operations also pollutes water and feed crops (Ibrahim *et al.*, 1997) [17]. Some authors studied the effect of cadmium on some freshwater bivalves including Ibrahim *et al.* (1997) [17] on *Caelatura aegyptiaca*, Saad and Emam (1998) [29] on *Caelatura teretiuscula* and Abdel Gawad (2005) [2] on *Caelatura prasidense*.

The present work aimed to assess the toxicity of Cd, as one of the most toxic metals frequently encountered in polluted areas, on *Lamellidens marginalis* and the ability of this freshwater bivalve to accumulate this toxic metal. This animal belongs to Bivalvia class of phylum Mollusca, which is second largest phylum (Verma and Prakash, 2020) [41].

Materials and Methods

After collection of the animals from habitat, they were immediately transported to the laboratory. The fouling and mud on shell valves were removed without disturbing the siphonal regions.

The equal sized animals (90-100 mm shell length) were grouped and kept in sufficient quantity of water (animal/liter) in aquaria with aeration for 24 Hrs. to adjust the animals in laboratory conditions (with renewal of water at interval of 12 to 13 hrs.). No food was given during acclimation time and during experiments. After 24 Hrs. 05 groups of animals of almost equal size (90-100 mm shell length) were formed and each group with 10 animals including control group and exposed to different test concentrations of cadmium for static bioassay tests. The stock solution of cadmium was prepared by was made dissolving appropriate quantity of cadmium chloride (CdCl₂ 2½H₂O AR Grade CDH Bombay) in double distilled water. The pH of the water is brought between 6.9 and 7.1 by adding 1N HCl (due to insolubility of cadmium in reservoir water having 7.6 to 8.1).

Results and Discussion

Table (1) shows the relation between the Cadmium concentration and the mortality rate of *Lamellidens marginalis*. The mortality rate did not exceed 10% during 96hrs at Cadmium concentrations of 1.5, 5.0 and 8.0 mg/l, in different season i.e., summer, monsoon and winter respectively while it was 100% at 6.0, 14.0 and 17.0 in summer, monsoon and winter respectively. The value of 96hrs LC50 of Cadmium on this species was found to be 3.0, 9.0 and 12.0 in summer, monsoon and winter respectively.

Table 1: 96-hour acute toxicity results of Cadmium on *Lamellidens marginalis* indifferent season.

| Concentration of Cd (mg/l) | | | Number of <i>L. marginalis</i> | Number of <i>L. marginalis</i> dead | Mortality rate (%) |
|----------------------------|---------|--------|--------------------------------|-------------------------------------|--------------------|
| Summer | Monsoon | Winter | | | |
| 1.5 | 5.0 | 8.0 | 10 | 1 | 10 |
| 2.0 | 6.0 | 9.0 | 10 | 2 | 20 |
| 2.5 | 7.0 | 10.0 | 10 | 3 | 30 |
| 3.0 | 8.0 | 11.0 | 10 | 4 | 40 |
| 3.0 | 9.0 | 12.0 | 10 | 5 | 50 |
| 4.0 | 10.0 | 13.0 | 10 | 6 | 60 |
| 4.5 | 11.0 | 14.0 | 10 | 7 | 70 |
| 5.0 | 12.0 | 15.0 | 10 | 8 | 80 |
| 5.5 | 13.0 | 16.0 | 10 | 9 | 90 |
| 6.0 | 14.0 | 17.0 | 10 | 10 | 100 |

Table 2: Bioaccumulation of Cd in soft tissue of *Lamellidens marginalis* after 96hrs acute toxicity to different concentrations of Cd solution.

| Concentration of Cd in solution (mg/l) | Concentration of Cd in soft tissue (mg/gm) |
|--|--|
| Experimental Control | 70.70 |
| 6.0 | 172.5 |
| 14.0 | 257.0 |
| 17.0 | 969.9 |

Bivalves are commonly available organisms that are abundant in the fresh water as well as in the marine environment. They have been suggested as ideal contamination indices in aquatic ecosystems because of their wide distribution, extensive population, sedimentary nature and the ability to accumulate contaminants (El Shenawy, 2004) [13]. Moreover, they close their shells for an extended period of time as escape behaviour or to exclude themselves from the outside environment when exposed to a contaminant (Wildridge *et al.*, 1998) [45]. Increasing use of contaminating chemicals in many industrialized parts of the world makes the development of

ecotoxicity measurement techniques is an absolute necessity (Brandao *et al.*, 1992) [9]. The first step is the acute toxicity test which can be carried out using bacteria, invertebrates and fish in order to show the potential risks of these chemicals (OECD, 1993; Yilmaz *et al.*, 2004) [23, 50].

The 96 hrs. LC50 of Copper and Zinc for the same species were 0.50 and 17.5 mg/l respectively (Abdel Gawad, 2001) [11] compared to 0.52 mg/l for Cd. This means that the toxicity of Cu > Cd > Zn and so this species is more tolerant to Zinc than to Cadmium or Copper. The differences between the values of LC50 of Cadmium for different bivalve species agree with Widerholm (1984) [44] and Voshell *et al.* (1989) [42], who stated that some marine organisms are very sensitive to heavy metals while others are resistant. In this study, the bioaccumulation of Cadmium in *Lamellidens marginalis* was increased by increasing the concentration of Cadmium solution, while the concentration factors decreased. This agrees with Wolf (1975) [46]; Kay (1985) [18]; Augier *et al.* (1992) [4]; Ramadan *et al.* (1997) [27] and Abdel Gawad (2005) [2].

The extent of contamination has been carried out to assess the status of pollutant concentrations in the water, sediment, and in organic tissue samples. The use of bioindicator species also provides additional information to survey environmental contaminations. Previous studies have determined the concentration of heavy metals (Beldi *et al.* 2006) [8] and investigated the impact of pollution on *D. trunculus* from the Gulf of Annaba (Amira *et al.* 2011; Soltani *et al.* 2012) [3, 38]. Moreover, the accumulation of heavy metals in *D. trunculus* is caused by human activities that affect the physiology of both genders of this species (Hamdani and Soltani-Mazouni 2011; Hamdani *et al.* 2014) [15, 16]. Indeed, high bioconcentration of heavy metal has been found in various Mollusk organisms, such as bivalves in many areas of the world (Neuberger-Cywiak *et al.* 2003; Sifi *et al.* 2007, 2013; Feldstein *et al.* 2003) [22, 34, 35, 14]. Bioconcentration is influenced by the physico-chemical properties of contaminants involved in the surrounding environmental conditions (e.g., biotope characteristics) and the biochemical/physiological adaptation of each organism (Barron 1990; Connell 1988) [6, 11]. Thus, compounds that accumulate in the tissues of aquatic organisms depend on several endogenous factors, namely species characteristics and trophic interactions (Mendez *et al.* 2001; Skinner *et al.* 2004) [21, 36], since Cd is a non-essential metal that is not physiologically present in organisms and can be toxic to humans, even at very low concentrations. In addition, this metal is relatively soluble, and tends to bioaccumulate rapidly by aquatic organisms such as bivalves (Shi *et al.* 2016) [32]. Cd is considered to be a highly toxic environmental pollutant and potent cell poison that causes different types of damage (Pascal *et al.* 2010) [24]. Therefore, the present study has shown that after treatment with LC50 and LC25 at 48 and 96 h, accumulation of Cd in *D. trunculus* increased in a concentration- and time-dependent manner. A significant increase in the tissue concentration appeared 48 h after exposure in planarians after treatment with 0.63 mg/l of Cd (Wu *et al.* 2012) [47]. In fact, the accumulation of Cd was also observed in other species exposed to metal contamination under laboratory conditions or in their natural environment such as bivalves *R. decussates* (Bebiano *et al.* 1993; Smaoui-Damak *et al.* 2003) [7, 37], *C. fluminea* (Legeay *et al.* 2005) [19], and *D. polymorpha* (Marie *et al.* 2006) [20], or in Gastropods like *Austrocochlea constricta* (Walsh *et al.* 1994) [43].

Acknowledgement

The authors express sincere thanks to the Capt. Dr. Maqdoom Farooqui Sir, Principal, Dr. Rafiq Zakira College for Women, Aurangabad for their valuable guidance.

References

1. Abdel Gawad SS. Studies on benthic invertebrates of Nile River at Helwan Region. Ph.D. Thesis, Fac. Sci., Mansoura Univ; c2001. p. 138.
2. Abdel Gawad SS. Toxicity and bioaccumulation of some heavy metals in fresh water bivalve (*Caelatura prasidens* Cailliaud, 1827). J Egypt. Acad. Soc. Environ. Develop. 2005;6(2):137-150.
3. Amira A, Sifi K, Soltani N. Measure of environmental stress biomarkers in *Donax trunculus* (Mollusca, Bivalvia) from the Gulf of Annaba (Algeria) Eur. J Exp. Biol. 2011;2:7-16.
4. Augier H, Park WK, Ramonda GR. Study of geographical seasonal metal content variations in different parts of the edible sea urchin *Paracentrotus lividus*, from three provincial test area. Revue internationale d, Oceanographie Medicale; c1992. p. 107-108:75-87.
5. Barak NAE, Mason CF. A catchment survey for heavy metals and using the eel *Anguilla anguilla* Chemosphere. 1990;21(4-5):695-699.
6. Barron MG. Bioconcentration (will water-borne organic chemicals accumulate in aquatic animals?). Environ Sci. Technol. 1990;24:1612-1618.
7. Bebianno MJ, Nott JA, Langston WJ. Cadmium metabolism in the clam *Ruditapes decussatus*: the role of metallothioneins. Aquat Toxicol. 1993;27:315-334.
8. Beldi H, Gimbert F, Maas S, Scheifler R, Soltani N. Seasonal variations of Cd, Cu, Pb and Zn in the edible mollusc *Donax trunculus* (Mollusca, Bivalvia) from the gulf of Annaba, Algeria. Afric. J Agric. Res. 2006;1(4):85-90.
9. Brandao C, Bohets HL, Vyver IE, Dierickx PJ. Correlation between the in vivo cytotoxicity to cultured fathead minnow fish cells and fish lethality data for 50 chemicals. Chemosphere. 1992;25:553-562.
10. Cicik B, Engin K. The effect of cadmium on levels of glucose in serum, glycogen reserves in the liver, muscle tissues of *Cyprinus carpio* (L. 1758). Turk J Vet Anim Sci. 2005;29:113-117.
11. Connell DW. Bioaccumulation behavior of persistent organic chemicals with aquatic organisms. Rev Environ Contam Toxicol. 1988;102:117-154.
12. Cross FA, Sunda WG. The relationship between chemical speciation and bioavailability of trace metals to marine organisms a review. In: Chao NL, Kirby-Smith W (Eds.) Proceedings of the symposium on utilization of coastal ecosystems, Rio Grande, RS-Brazil. 1985;1:169-182.
13. El Shenaway NS. Heavy metal and microbial depuration of the clam *Ruditapes decussates* and its effect on bivalve behavior and physiology. Environ. Toxicol. 2004;19:143-153.
14. Feldstein T, Kashman Y, Abelson A, Fishelson L, Mokaday O, Bresler V, et al. Marine molluscs in environmental monitoring. III trace metals and organic pollutants in animal tissue and sediment. Helgoland Mar Res. 2003;57:212-219.
15. Hamdani A, Soltani Mazouni N. Changes in biochemical composition of the gonads of *Donax trunculus* L. (Mollusca, Bivalvia) from the Gulf of Annaba (Algeria) in relation to reproductive events and pollution. Jordan J Biol. Sci. 2011;4:149-156.
16. Hamdani A, Soltani Mazouni N, Soltani N. Quantitative and qualitative analysis of proteins in gonads of *Donax trunculus* from the Annaba Bay: effects of site, season and sex Adv. Environ. Biol. 2014;8:740-749.
17. Ibrahim AA, Sleem SH, Bahgat FG, Ali AS. Effect of certain water pollutants on the biology of the fresh water clam *Caelatura (unio) aegyptiaca* (Bivalvia). Egypt. J Aquat. Biol. & Fish. 1997;1:47-65
18. Kay SH. Cadmium in aquatic food webs. Residue, Rev. 1985;96:13-43.
19. Legeay A, Achard-Joris M, Baudrimont M, Massabuau JC, Bour-dineaud JP. Impact of cadmium contamination and oxygen levels on biochemical responses in the Asiatic clam *Corbicula fluminea*. Aquat Toxicol. 2005;74:242-253.
20. Marie V, Baudrimont M, Boudou A. Cadmium and zinc bioaccumulation and metallothionein response in two freshwater bivalves (*Corbicula fluminea* and *Dreissena polymorpha*) transplanted along a polymetallic gradient. Chemosphere. 2006;65:609-617.
21. Mendez E, Giudice H, Pereira A, Inocente G, Medina D. Total mercury content fish weight relationship in swordfish (*Xiphias gladius*) caught in the southwest Atlantic Ocean. J Food Compos Anal. 2001;14:453-460.
22. Neuberger-Cywiak L, Achituv Y, Garcia EM. Effects of zinc and cadmium on the burrowing behavior LC50 and LT50 on *Donax trunculus* Linnaeus (Bivalvia-Donacidae). Bull Environ Contam Toxicol. 2003;70:713-722.
23. OECD (Organization for Economic Co-operation and Development): OECD Guidelines for testing of chemicals. OECD Paris; c1993.
24. Pascal PY, Fleeger JW, Galvez F, Carman KR. The toxicological interaction between ocean acidity and metals in coastal meiobenthic copepods. Mar Pollut Bull. 2010;60:2201-2208.
25. Rainbow PS. The biology of heavy metals in the sea. Int. J Environ Stud. 1985;25:195-211.
26. Mukhtar M, Muhammad H, Ibrahim S, Ahmad G, Muhammad Y, Abubakar S. Antioxidant and acute toxicity of stem extracts of the *Ficus iteophylla*. Int. J Adv. Chem. Res. 2020;2(1):17-19. DOI: 10.33545/26646781.2020.v2.i1a.18
27. Ramadan SHE, Khalil AN, Dowidar NM, El Sonbaty SF. Toxicity and bioaccumulation of Cu and Zn in two isopod species (Crustacea Bull. Nat. Inst. Oceanogr. & Fish. A.R.E. 1997;23:135-154.
28. Roméo M, Frasila C, Gnassia-Barelli M, Damiens G, Micu D, Mus-tata G. Biomonitoring of trace metals in the Black Sea (Romania) using mussels *Mytilus galloprovincialis*. Water Res. 2005;39(4):596-604.
29. Saad AA, Emam WM. Toxicity and bioaccumulation of heavy metals in the fresh water bivalve *Calatura teretiuscula* Egypt. J Biol. and fish. 1998;2(4):15-27.
30. Sanders MJ. A field evaluation of the freshwater river crab, *Potamonautes warreni*, as a bio-accumulative indicator of metal pollution. Rand Afrikaans University, South Africa; c1997.
31. Selvi M, Gul A, Yilmaz M. Investigation of acute toxicity of Cadmium Chloride (CdCl₂ H₂O) metal salt

- and behavioral changes it cause on in water Frog (*Rana ridibunda* palls, 1771). Chemosphere. 2003;52:259-263.
32. Shi W, Zhao X, Han Y, Che Z, Chai X, Liu G. Ocean acidification increases cadmium accumulation in marine bivalves: a potential threat to seafood safety. Sci. Rep. 2016;6:20197. <https://doi.org/10.1038/srep20197>.
 33. Shinde NG. Seasonal changes in the oocytes of Freshwater Mollusc, *Lamellidens corrianus* (Lea). International Journal of Biological Innovations. 2021;3(1):199-204. <https://doi.org/10.46505/IJBI.2021.3121>
 34. Sifi K, Amira A, Soltani N. Oxidative stress and biochemical composition in *Donax trunculus* (Mollusca, Bivalvia) from the gulf of Annaba (Algeria). Adv. Environ Biol. 2013;7(4):595-604.
 35. Sifi K, Chouahda S, Soltani N. Biosurveillance del'environnement par la mesure de biomarqueurs chez *Donax trunculus* dans le golfe d'Annaba (Algérie). Mésogée. 2007;63:11-18.
 36. Skinner C, Turoczy NJ, Jones PL, Barnett D, Hodges R. Heavy metal concentrations in wild and cultured Black lip Abalone (*Haliotis rubra* Leach) from southern Australian waters. Food Chemistry. 2004;85:351-356.
 37. Smaoui-Damak W, Hamza-Chaffai A, Berthet B, Amiard JC. Preliminary study of the clam *Ruditapes decussates* exposed in situ to metal contamination and originating from the Gulf of Gabès, Tunisia. Bull Environ Contam Toxicol. 2003;71:961-970.
 38. Soltani N, Amira A, Sifi K, Beldi H. Environmental monitoring of the Annaba gulf (Algeria): measurement of biomarkers in *Donax trunculus* and metallic pollution ecotoxicology Bull. Soc. Zool. Fr. 2012;137:47-56.
 39. Sthanadar IA, Sthanadar AA, Begum B, Nair MJ, Ahmad I, Muham-mad A, et al. Aquatic pollution assessment using skin tissues of Mulley (*Wallago attu*, Bloch & Schneider, 1801) as a bio-indicator in Kalpani River at District Mardan, Khy-ber Pakhtunkhwa. J Biodivers Environ Sci. 2015;6:57-6.
 40. Ukoha PO, Ekere NR, Udeogu UV, Agbazue VE. Potential health risk assessment of heavy metals [Cd, 17 Cu and Fe] concentrations in some imported frozen fish species consumed in Nigeria. Int. J Chem. Sci. 2014;12:366-374.
 41. Verma AK, Prakash S. Status of Animal Phyla in different Kingdom Systems of Biological Classification. International Journal of Biological Innovations. 2020;2(2):149-154. <https://doi.org/10.46505/IJBI.2020.2211>
 42. Voshell JR, Layton JRRJ, Hiner SW. Field techniques for determining the effects of toxic substances on benthic macro invertebrates in rock-bottom streams. In Aquatic Toxicology and Hazard Assessment. 12 ASTMSTP1027 Cowgill, U.M. and Williams, L.R. (Ed) American society for testing and materials. Philadelphia, PA; c1989. p. 134-155.
 43. Walsh K, Dunstan RH, Murdoch RN, Conroy BA, Roberts TK, Lake P. Bioaccumulation of pollutants and changes in population parameters in the gastropod mollusk, *Austrocochlea constricta*. Arch Environ Contam Toxicol. 1994;26:367-373.
 44. Wiederholm T. Response of aquatic insects to environmental pollution In: The ecology and Aquatic Insects. Res, V.H. and Rosenberg, D.M. (Ed). Proger Publishers New York, N.Y; c1984. p.508-557.
 45. Wildridge PL, Werner RG, Doherty FG, Neuhauser EF. Acute effects of potassium on filtration rates of adult zebra mussels *Dreissena polymorpha*. J Great Lakes Res. 1998;24:629-636.
 46. Wolf PD. Mercury content of mussels from west European coast. Mar. Pollut. Bull. 1975;6:61-23.
 47. Wu JP, Chen HC, Li MH. Bioaccumulation and toxicodynamics of cadmium to freshwater Planarian and the protective effect of N-acetylcysteine. Arch Environ Contam Toxicol; c2012. <https://doi.org/10.1007/s00244-012-9764-5>.
 48. Yasmeen S. Cadmium induced histopathological alterations in female gonad of freshwater bivalve mollusks, *Lamellidens marginalis* during summer season. International Journal of Biological Innovations. 2019;1(2):73-77. <https://doi.org/10.46505/IJBI.2019.1207>
 49. Yasmeen S, Pathan TS, Pawar RT. Effect of cadmium chloride on glycogen level in some organs of freshwater bivalve, *Lamellidens marginalis*. International Journal of Biological Innovations. 2021;3(2):367-372. <https://doi.org/10.46505/IJBI.2021.3218>
 50. Yilmaz M, Gul A, Karakose E. Investigation of acute toxicity and the effect of Cadmium Chloride ($CdCl_2 \cdot H_2O$) metal salt on behavior of guppy (*Poecilia reticulata*) Chemosphere. 2004;56:375-380.
 51. Rainbow PS, White SL. Comparative strategies of heavy metal accumulation by crustaceans: zinc, copper and cadmium in a decapod, an amphipod and a barnacle. Hydrobiologia. 1989;174:245-262.